

Evaluation of Antifungal and Phytochemical Activity from Different Parts of *Cerbera odollam* **Gaertn**

by

Chu Sue Yin (1331111136)

A thesis submitted in fulfillment of the requirements for the degree of Master of Science in Bioprocess Engineering

School of Bioprocess Engineering UNIVERSITI MALAYSIA PERLIS

ACKNOWLEDGMENT

Sincere gratitude is hereby extended to the following that never ceased in helping until paper is completed:

Assoc. Prof. Dr. Muhammad Syarhabil Ahmad, main supervisor and thesis advisor, for the unwavering guidance;

Assoc. Prof. Dr. Harbant Singh, co-supervisor, for sharing and instructing in thesis writing;

Prof. Dr. Awang Soh, co-supervisor for advices in laboratory;

Fundamental Research Grant Scheme (FRGS) (9003-00372) for financial support;

The members of panels for sharing their precious time and positive insights;

The PLVs and technicians who were willing to help when conducting experiment in laboratory;

My family and friends for moral support.

Chu Sue Yin (1331111136)

TABLE OF CONTENTS

THE	SIS DECLARATION	PAGE i
ACK	NOWLEDGMENT	ii
TAB	LE OF CONTENTS	iii
LIST	T OF TABLES	vi
LIST	COF FIGURES COF ABBREVIATIONS TRAK TRACT PTER 1 INTRODUCTION Background of study Problem Statement Objective 1.3.1 Overall Goal 1.3.2 Specific Objectives	viii
LIST	OF ABBREVIATIONS	xii
ABS	ГРАК	xiii
ABS	TRACT	xiv
СНА	PTER 1 INTRODUCTION	
1.1	Background of study	1
1.2	Problem Statement	3
1.3	Objective	4
	1.3.1 Overall Goal	4
	1.3.2 Specific Objectives	4
1.4	Scope of Study	5
	.,	
СНА	PTER 2 LITERATURE REVIEW	
2.1	Introduction	6
2.2	Cerbera odollam Gaertn	
	2.2.1 Botanical Description and Localization	6
	2.2.2 Bio-compounds in Cerbera odollam	8
	2.2.3 Applications of Cerbera odollam extracts	11
2.3	Post harvest fungi	15
2.4	Fungicides	17
2.5	Extraction technique	19
2.6	Antifungal susceptible testing	22
2.7	Toxicology test	23

2.8	Liquio	d Chromatography Mass Spectrophotometer (LCMS)	24
СНА	PTER 3	3 METHODOLOGY	
3.1	Introd	luction	27
3.2	Chem	icals	28
3.3	Plant	materials collection	28
3.4	Plants	extraction	30
3.5	Funga	al strain and suspensions preparation	31
3.6	Antifu	ungal assay	
	3.6.1	Preparation of extract concentration	32
	3.6.2	Preparation of modified potato dextrose agar (PDA)	34
	3.6.3	Antifungal test	34
3.7	Deter	mination o f minimum inhibition concentration (MIC)	35
3.8	Brine	shrimp test (BST) bioassay	
	3.8.1	Preparation of extracts concentrations	37
	3.8.2	Preparation of artificial sea water and shrimp	37
	3.8.3	Brine shrimp lethality bioassay	38
3.9	Phyto	chemical analysis	
	3.9.1	Test for alkaloids	40
	3.9.2	Test for anthraquinone	40
	3.9.3	Test for cardiac glycoside	40
	3.9.4	Test for flavonoid	41
	3.9.5	Test for phenol	41
	3.9.6	Test for saponin	41
(3.9.7	Test for steroid	42
	3.9.8	Test for tannin	42
	3.9.9	Test for terpenoid	42
3.10	Liquio	d Chromatogram Mass Spectrophotometer (LCMS)	43
СНА	PTER 4	4 RESULT AND DISCUSSIONS	
4.1	Characte	ristic of samples	44
4.2 E	Extractio	on	46
4.3 P	hytoche	emical analysis	47

4.4	Antifunga	al bioassay	
	4.4.1	Effect of plant extracts on the growth of Aspergillus niger	49
	4.4.2	Effect of plant extracts on the growth of Penicilium citrinum	52
	4.4.3	Effect of plant extracts on the growth of Fusarium oxysporum	55
4.5	Minimum	n fungi inhibition	58
4.6	Toxicity t	test	60
4.7	Liquid ch	romatography mass spectrum analysis	
	4.7.1	Leaf extracts analysis	66
	4.7.2	Seed extracts analysis	71
	4.7.3	Fruit extracts analysis	73
	4.7.4	Flower extracts analysis	77
	4.7.5	Fruit extracts analysis Flower extracts analysis Bark extracts analysis Wood extracts analysis Summary of LCMS extracts analysis S CONCLUSION On	81
	4.7.6	Wood extracts analysis	87
	4.7.7	Summary of LCMS extracts analysis	88
CH	APTER 5	CONCLUSION	
5.1	Conclusio	on	92
5.2	Recomme	endation	94
		XO CONTRACTOR OF THE PROPERTY	
RE	FERENC	ES	95
		.6	
AP	PENDIX .	Summary of LCMS extracts analysis CONCLUSION on endation ES A BLICATIONS	104
	. (
LIS	T OF PU	BLICATIONS	116
LIS	T OF AV	VARDS	117

LIST OF TABLES

NO.		PAGE
2.1	Summary of bio-compounds from different part of <i>Cerbera odollam</i> (leaf, seed, stem, bark and latex).	10
2.2	Summary of the tested arena and application for different parts (seed, bark, leaf, fruit, wood, flower and root) of <i>Cerbera odollam</i> .	12
2.3	List of fungi and disease associated with fruits and vegetables.	16
2.4	Some of the commercial bio-fungicide in the market and the specific target organism	19
2.5	Summary of the experimental conditions (type of solvent, techniques and extraction times) for different parts of <i>Cerbera odollam</i> .	21
2.6	Interpretative range for fungal inhibition activity according to classes and inhibition value (%).	22
2.7	Toxicity category based on the route of exposure (by oral, dermal or inhalation) applied to tested organism.	24
3.1	Chemicals and the brands.	28
4.1	Phytochemical constituents of leaf, seed, fruit, flower, bark and wood's crude extract of <i>Cerbera odollam</i> . The tests were carried out under standard procedure.	48
4.2	Colony diameter (cm) of <i>Aspergillus niger</i> after undergone treatment with six extraction parts of <i>Cerbera odollam</i> at four different concentrations (500, 1000, 2000 and 3000 ppm) for seven days at 25 \pm 2 °C.	51
4.3	Colony diameter (cm) of <i>Penicilium citrinum</i> after undergone treatment with six extraction parts of <i>Cerbera odollam</i> at four different concentrations (500, 1000, 2000 and 3000 ppm) for seven days at 25 \pm 2 °C.	54
4.4	Colony diameter (cm) of <i>Fusarium oxysprorum</i> after undergone treatment with six extraction parts of <i>Cerbera odollam</i> at four different concentrations (500, 1000, 2000 and 3000 ppm) for seven days at 25 \pm 2 \times	57
4.5	Minimum inhibition concentration (ppm) of six extracts (leaf, seed, fruit, flower, bark and wood) on <i>Aspergillus niger</i> , <i>Fusarium oxysporum</i> and <i>Penicilium citrinum</i> after seven days incubation at 25 \pm 2 °C. The test was performed at sterile 96-well plates.	59

- 4.6 Mortality (%) of nauplii in Brine Shrimp Test (BST) of six plant extracts at nine different concentrations (5 ppm to 1280 ppm). [Mortality (%) was calculated as percentage of nauplii survives for control minus percentage of nauplii survives for treatment].
- 4.7 Summarized of LC₅₀ value (ppm) in each extracts and the category of toxicity. [Toxicity category: Highly toxic = 0 to 0.2 mg/L; Moderate toxic = 0.2 to 2 mg/L; Slightly toxic = 2 to 20 mg/L, relatively non toxic = more than 20 mg/L (Damalas & Eleftherohorinos, 2011)]
- 4.8 Summary of compounds found in six extract of Cerbera odollam (leaf, seed, fruit, flower, bark and wood) with corresponding molecular weight (MW).

 89 seed, fruit, flower, bark and wood) with corresponding molecular weight (MW).

LIST OF FIGURES

NO. 2.1	(A) flowers (B) fruits (C) tree of <i>Cerbera odollam</i> .	PAGE
2.2	Compounds structure of: A, Cerberalignan (lignan);	8
2.2	B, Cerberin (cardiac glycoside) (Shen <i>et al.</i> , 2007)	0
2.3	(i) Conidia head of <i>Aspergillus</i> spp. (ii) <i>Penicilli</i> (iii) <i>Fusarium</i> spp. (Hussain <i>et al.</i> , 2012; Lhan <i>et al.</i> , 2006)	16
2.4	Basic components of LCMS system (Sargant, 2013)	25
3.1	Research methodology flow chart.	27
3.2	Preparation samples of <i>Cerbera odollam</i> (flower, leaf, wood, bark, fruit and seed).	29
3.3	(a) Preparation of samples extracts (leaf, flower, fruit, seed, bark and wood) by soxhlet extraction method; (b) concentration of filtrate using rotary evaporator at 40 $^{\circ}$ C; (c) color of each crude extracts.	31
3.4	Spores of (a) Aspergillus niger, (b) Fusarium oxysporum and (c) Penicilium citrinum under microscope (x100) after serial of dilution to obtain spores in $1x10^7$ spore/mL.	32
3.5	Concentrations of (a) leaf and (b) bark extracts from 3000 to 500 ppm (right to left) through serial of dilution.	33
3.6	Sterilized 96-well plates were used to perform the Minimum Inhibition Concentrations (MIC) for the tested fungi. A total of 8 rows and 12 columns for the plate.	36
3.7	Setup for breeding the shrimp inside the beaker with autoclaved water. Continuous air and light were supplied for the whole process. The breeding process was performed under laminar flow for 24 hours.	38
3.8	The Dionex-Ultimate ® 3000 Rapid Separation LC system (connected with a Micro TOF-Q mass spectrometer.	43
4.1	The (a) young fruit and (b) mature fruit of <i>Cerbera odollam</i> collected from Pengkalam Asam, Perlis, Malaysia.	44
4.2	The moisture contents of six different parts of Cerbera odollam at $103~\mathrm{C}$ for 24 hours.	45
4.3	The extraction yield of six different parts of <i>Cerbera odollam</i> after 8 hour of extraction using soxhlet extractor.	46

- 50 4.4 Inhibition zone (%) of six extraction parts of *Cerbera odollam* extracts on Aspergillus niger at 25±2 °C for seven days inside the incubator with four different concentrations (500 to 3000ppm) applied. 4.5 Fungi growth observed on potatoes dextrose agar (PDA) medium 51 treated with extract (500 ppm and 3000 ppm) after seven days incubation at 25±2 °C. The pictures showed the decreasing trend of inhibition diameter of flower extracts on Aspergillus niger at 500 ppm and 3000 ppm. 4.6 Inhibition zone (%) of six parts of Cerbera odollam extracts on 53 Penicilium citrinum at 25 ± 2 °C for seven days inside the incubator with four different concentrations (500 to 3000ppm) applied. 4.7 Fungi growth observed on potatoes dextrose agar (PDA) medium 55 treated with extract (500 ppm and 3000 ppm) after seven days incubation at 25 ± 2 °C. The pictures showed the decreasing trend of inhibition diameter of wood extracts on *Penicilium citrinum* at 500 ppm and 3000 ppm. Inhibition zone (%) of six parts of Cerbera odollam on Fusarium 4.8 56 oxysprorum at 25 ± 2 °C for seven days inside the incubator incubator at four different concentrations (500 to 3000ppm) applied. 4.9 Fungi growth observed on potatoes dextrose agar (PDA) medium 57 treated with extract (500 ppm and 3000 ppm) after seven days incubation at 25±2 °C. The pictures showed the decreasing trend of inhibition diameter of wood extracts on Fusarium oxysporum at 500 ppm and 3000 ppm. 4.10 Mortality response of nauplii which exposed to different concentration 61 (5 to 1280 ppm) of leaf extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm.
- 4.11 Mortality response of nauplii which exposed to different concentration (5 to 1280 ppm) of seed extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm.
- 4.12 Mortality response of nauplii which exposed to different concentration (5 to 1280 ppm) of fruit extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm.
- 4.13 Mortality response of nauplii which exposed to different concentration (5 to 1280 ppm) of flower extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm.

4.14 Mortality response of nauplii which exposed to different concentration 63 (5 to 1280 ppm) of bark extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm. Mortality response of nauplii which exposed to different concentration 64 (5 to 1280 ppm) of wood extracts for 24 hours under laminar flow. The concentration of extracts which kill 50% of nauplii was calculated from the linear equations of the graph, by taking the antilogarithm. LCMS Chromatogram of leaf extract of Cerbera odollam at UV 254 66 4.16 nm and the compounds labeled were further examine under ESI-MS. 4.17 LCMS total ion chromatograms of detected compounds in leaf extract 67 acquired at ESI(+) and ESI(-) (i) The MS/MS spectrum of (a) at m/z 191, t_R 10.3 min; (ii) MS/MS 4.18 68 spectrum of carbendazim in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the carbendazim existence in the leaf extract. 4.19 (i) The MS/MS spectrum of (b) at m/z 138, t_R 29.7 min; (ii) MS/MS 69 spectrum of hydroxybenzoic acid in mass bank (Anonymous, 2015). The both MS/MS spectrum matched affirm the hydroxybenzoic acid existence in the leaf extract. (i) The MS/MS spectrum of (c) at m/z 534, t_R 38 min; (ii) MS/MS 4.20 70 spectrum of neriifolin in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the neriifolin existence in the leaf extract. LCMS Chromatogram of seed extract of Cerbera odollam at UV 254 71 4.21 nm and the compounds labeled were further examine under ESI-MS. LCMS sum ion chromatograms of detected compounds in seed extracts 4.22 72 acquired at ESI(+) and ESI(-) 4.23 LCMS Chromatogram of fruit extract of Cerbera odollam at UV 254 73 nm and the compounds labeled were further examine under ESI-MS. LCMS sum ion chromatograms of detected compounds in fruit extracts 4.24 74 acquired at ESI(+) and ESI(-) (i) The MS/MS spectrum of (d) at m/z 306, t_R 21,4 min; (ii) MS/MS 75 4.25 spectrum of epigallocatechin in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the epigallocatechin existence in the fruit extract.

76 4.26 (i) The MS/MS spectrum of (e) at m/z 290, t_R 29.2 min; (ii) MS/MS spectrum of epicatechin in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the epicatechin existence in the fruit extract. 4.27 (i) The MS/MS spectrum of (f) at m/z 303, t_R 32.1 min; (ii) MS/MS 77 spectrum of quercetin in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the quercetin existence in the fruit extract. 4.28 LCMS Chromatogram of flower extract of Cerbera odollam at UV 254 78 nm and the compounds labeled were further examine under ESI-MS. LCMS sum ion chromatograms of detected compounds in flower 79 4.29 extracts acquired at ESI(+) and ESI(-). (i) The MS/MS spectrum of (g) at m/z 740, t_R 30.3 min; (ii) MS/MS 80 4.30 spectrum of kaempferol- 3-galactoside-6"-rhamnoside-3"'-rhamnoside in mass bank (Anonymous, 2015). The both MS/MS spectrum matched affirm the kaempferol existence in the flower extract. LCMS Chromatogram of bark extract of Cerbera odollam at UV 254 81 nm and the compounds labeled were further examine under ESI-MS. 4.32 LCMS total ion chromatograms of detected compounds in bark extracts 82 acquired at ESI(+) and ESI(-). The MS/MS spectrum of (i) at m/z 191, t_R 25.8 min for cerberinic acid 4.33 83 and its structures in bark extracts. (i) The MS/MS spectrum of (j) at m/z 205, t_R 27.2 min; (ii) MS/MS 4.34 84 spectrum of cerbinal in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the cerbinal existence in the bark extract. (i) The MS/MS spectrum of (k) at m/z 137, t_R 34.3 min; (ii) MS/MS 85 spectrum of salicylic acid in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the salicylic acid existence in the bark extract. (i) The MS/MS spectrum of (l) at m/z 353, t_R 24 min; (ii) MS/MS 86 spectrum of terephthalic acid in mass bank (Anonymous, 2015). Both the MS/MS spectrum matched affirm the terephthalic acid existence in the bark extract. LCMS Chromatogram of wood extract of Cerbera odollam at UV 254 4.37 87 nm and the compounds labeled were further examine under ESI-MS. LCMS total ion chromatograms of detected compounds in wood 88 4.38 extracts acquired at ESI(+) and ESI(-).

LIST OF ABBREVIATIONS

Brine Shrimp Test BST

DFB Diflubenzuron

Electron Spray Ionization ESI

FCX

Liquid Chromatography Mass Spectrometry

Minimum Inhibition Concentration

Potato Dextrose Agar

Potato Dextrose Broth

Total Ion Current **LCMS**

MIC

PDA

PDB

TIC

Thin Layer Chromatograhy TLC

Nuclear Magnetic Resonance NMR OThis item is

Penilaian Aktiviti Antikulat dan Fitokimia daripada Bahagian-bahagian Berbeza Cerbera Odollam Gaertn

ABSTRAK

Penggunaan racun kulat kommercial berlebihan dalam sektor pertanian telah mengakibatkan pencemaran alam sekitar yang akan membahayakan kesihatan manusia. Kajian ini bertujuan untuk meneroka kegunaan ekstrak tanaman (botani) sebagai pengganti kepada sintetik racun kulat. Ekstrak Etanol daripada bahagian tumbuhan Cerbera odollam seperti daun, bunga, buah, biji benih, kayu, dan kulit kayu telah diuji bagi menentukan sifat antikulat. Bioesei antikulat telah dijalankan dengan menggunakan kaedah pencairan dengan kepekatan yang berlainan (500 ke 3000 ppm) terhadap kulat berikut: Aspergillus niger, Fusarium oxysporum and Penicilium citrinum dan seterusnya dinilai dengan Minimum Inhibitory Concentration (MIC). Toksikologi tanaman dikaji dengan kaedah Brine Shrimp Test (BST) di mana kepekatan yang berbeza digunakan (5 ke 1280 ppm). Kajian fitokimia pula telah dijalankan melalui prosedur piawai untuk mengetahui sebatian fitokimia yang terlibat dalam aktiviti antikulat. Kaedah *Liquid Chromatography Mass Spectrometry* (LCMS) telah dijalankan untuk menentukan sebatian antikulat yang wujud dalam ekstrak tanaman. Keputusan kajian secara umumnya telah menunjukkan zon inhibition terhadap kulat di semua bahagian yang dikaji. Antara bahagian yang dikaji, daun menunjukkan kesan antikulat yang penting (P \leq 0.05) terhadap Aspergillus niger and Fusarium oxysporum dalam semua kepekatan (500 ke 3000 ppm) di mana 3000 ppm adalah terbaik antaranya. Walau bagaimanapun, tiada kesan ketara apabila ekstrak diuji ke atas Penicilium citrinum. Dalam kajian MIC, dos yang paling rendah dari daun dan kulit kayu adalah 250 ppm terhadap Aspergillus niger manakala biji benih, buah, bunga dan kayu menunjukkan 500 ppm; *Penicilium citrinum* and *Fusarium oxysporum* merekodkan 500 ppm untuk semua ekstrak. Bagi kajian toksikologi, nilai kritikal selamat ialah 20 ppm ke atas dan 2 ppm ke bawah adalah tidak selamat untuk manusia. Kayu (5619.97 ppm), buah (2116.66 ppm), kulit kayu (1745.04 ppm) dan bunga (64.47 ppm) selamat diggunakan manakala tahap selamat bagi daun (8.31 ppm) dan biji benih (3.62 ppm) adalah sedikit toksik dan harus digunakan dengan waspada. Kajian LCMS daripada ekstrak bunga dan buah Cerbera odollam adalah yang pertama. Sebatian antikulat yang telah dikesan ialah neriifolin, asid hidroksibenzoik, asid cerberinik, asid salisilik dan asid terepthtalik. Neriifolin dan asid hidroksibenzoik merupakan sebatian yang pertama kali dikenal pasti dalam ektrak buah, bunga, dan kayu; asid cerberinik, asid salisilik dan asid terepthtalik dalam ekstrak kulit kayu. Kajian ini menunjukkan daun Cerbera odollam mempunyai potensi untuk dijadikan racun kulat bio.

Evaluation of Antifungal and Phytochemical Activity from Different Parts of Cerbera Odollam Gaertn

ABSTRACT

Heavy usage of commercial fungicide in the agricultural sector has resulted in environmental pollution that has imposed a significant risk to human health. The current research intended to explore the use of plant extracts (botanical) as alternative to synthetic fungicide. Ethanolic extracts of Cerbera odollam from leaf, flower, fruit, seed, wood and bark were tested for antifungal properties. Antifungal bioassay was performed through dilution method at various concentrations (500 ppm to 3000 ppm) against fungi: Aspergillus niger, Fusarium oxysporum and Penicilium citrinum and assessed based on the Minimum Inhibitory Concentration (MIC). Plant toxicity was tested by Brine Shrimp Test (BST) at different concentrations (5 to 1280 ppm). Phytochemical tests were done using standard procedures to identify the phytochemical compounds involved in the antifungal activity and Liquid Chromatography Mass Spectrometry (LCMS) was performed to determine possible antifungal compounds existed in all extracts. The results of the research showed that the inhibition zone for the tested fungi generally exhibited antifungal activity from different parts. Among the treatments, leaf extracts had recorded significant antifungal effects (P \leq 0.05) against Aspergillus niger and Fusarium oxysporum at all concentrations (500 to 3000 ppm) with 3000 ppm showing the best inhibition zone. However, there was no significant difference when extracts were tested against *Penicilium citrinum*. For the MIC study, the lowest dosage recorded for leaf and bark was 250 ppm against Aspergillus niger, while seed, fruit flower and wood showed the effect at 500 ppm; Penicilium citrinum and Fusarium oxysporum recorded MIC value at 500 ppm from all treatments. In toxicology test, the safe value is an amount over 20 ppm while 2 ppm and below is considered unsafe or toxic for human being. Wood (5619.97 ppm), fruit (2116.66 ppm), bark (1745.04 ppm) and flower (64.47 ppm) extracts showed safe levels while leaf (8.31 ppm) and seed (3.62 ppm) were slightly toxic and should be used with caution. LCMS study of Cerbera odollam's flower and fruit crude extracts were first reported in the research. Antifungal compounds detected were neriifolin, hydroxybenzoic acid, cerberinic acid, salicylic acid and terephtalic acid. Neriifolin and hydroxybenzoic acid were identified for the first time from fruit, flower and wood extracts and cerberinic acid, salicylic acid and terephtalic acid from bark extracts. The results of current study indicated that *Cerbera odollam* leaf extracts has a potential of being a biofungicide.

CHAPTER 1

INTRODUCTION

1.1 Background of study

Post harvest management in agricultural production has been growing considerably over the past four decades due to the growing populations across the Asia-pacific region. In 2004, global production for fruits and vegetables in the Asia-pacific regions was approximated between 31% and 42%. However, due to diseases, poor handling technique and management, the losses in postharvest was reported between 24% to 40% (Asian Productivity Organization, 2006).

Post harvest diseases are mainly caused by fungi and bacteria. The fungi responsible for most post harvest disease are *Fusarium*, *Alternaria* and *Penicilium* (Coates & Johnson, 2013). In order to overcome the problem, commercial fungicide such as Imazalil, Benomyl, and Thiabenzole are extensively applied for postharvest disease control in fruits and vegetables (Singh *et al.*, 2011). Some of the fungicide are reported to be hazardous to human health, include vinclozolin, flucycloxuron (FCX) and diflubenzuron (DFB) (Rouabhi, 2010). These fungicides are not only able to have an effect on natural non-target organisms, but also accumulate as residues in food for human consumption (Okwute, 2012). Negative effects of synthetic fungicide are therefore raising grave concern globally.

Over the past 150 years, botanical pesticides have been traditionally used in the agricultural community. Anti-microbial activity of botanical has been reported by many researchers in their work (Ashikur *et al.*, 2011; Cowan, 1999; Duke *et al.*, 2010; Ghassan *et al.*, 2012). However, due to the regulatory procedures associated with product development, there are only a small number of commercial botanical pesticide available in the market today, such as *pyrethrins*, *rotenone*, *neem*, *ryania*, *nicotin*, *sabadilla*, *garlic oil* and *Capsicum* oleoresin (Okwute, 2012). Extract from natural plant products are preferred for replacing synthetic fungicide because of safe usage, as they are biodegradable.

Cerbera odollam Gaertn belonging to Apocynaceae family is commonly known as "Pong-Pong tree". It is also known as "Buta-buta" in Malay. In peninsular Malaysia, locations of Cerbera odollam are distributed mostly at coastal mangrove swamps, sometimes inland as a roadside tree at Perlis, Kedah, Perak, Selangor, Melaka, Pahang and Johor. This species is able to produce flowers and fruits throughout the year (Kiew et al., 2011).

In the past decades, *Cerbera odollam* has been widely studied on its effects as antiproliferative, anticancer, antiestrogenic, antimicrobial, antinociceptive and sedative effect at different dosage (Ahmed *et al.*, 2008; Ahmed *et al.*, 2006; Shen *et al.*, 2007). The bio-compounds reported in *Cerbera* includes cardiac glycoside, lignans, terpenoids, flavonoids, and some other compounds (Shen *et al.*, 2007; Yamauchi *et al.*,1987). Secondary metabolites which have the potential to defend against pathogens are classified into three main chemical constituent family, i.e. alkaloids, terpenoids and phenolic (Brusotti *et al.*, 2014). These secondary metabolite compounds have indicated their anti-microbial effects (Cowan, 1999; Cunha *et al.*, 2012; Soković *et al.*, 2009).

Thus, potential of *Cerbera odollam* as a replacement to conventional fungicide in post harvest management led the author to pursue in the anti-fungal effect in postharvest management.

In this research, the focus was on the evaluation of antifungal properties of different parts from *Cerbera odollam*, i.e. leaf, flower, fruit, seed, wood and bark. Ethanolic extracts of the plant parts were investigated for their antifungal activities and the bio-compounds through Liquid Chromatography Mass Spectrometer (LCMS).

1.2 Problem Statement

Cerbera odollam has been widely studied on its antimicrobial or anticancer in medical application. However, little investigation has been conducted in inhibiting agricultural diseases. Roles of this species in agro industry are still in the infancy stage. Therefore, further study could be done in this area. Besides, up to date, there are no reports available regarding the bio-compounds obtained from Cerbera odollam flowers and fruits. Hence, further analysis of these plant parts could be done by using LCMS.

1.3 Objective

1.3.1 Overall Goal

The aim of this research is to evaluate *Cerbera odollam* extracts as an alternative to commercial fungicide in post harvest management.

1.3.2 Specific Objectives

The objectives of this research are:

- i. To evaluate the antifungal effect from different parts of *Cerbera odollam*, such as leaf, flower, fruit, seed, wood and bark on the growth of *Aspergillus niger*, *Penicilium citrum* and *Fusarium oxysporum*.
- ii. To evaluate the toxicity (LC $_{50}$) of the plant extracts using Brine Shrimp Test (BST).

iii. To determine the correlation between phytochemical compounds and antifungal activity from different parts of plant extracts.

1.4 Scope of Study

Aspergillus niger, Fusarium oxysporum and Penicilium citrum are common postharvest pathogens. The scope of the study is to investigate the anti-fungal activity from different parts of crude extracts of Cerbera odollam e.g leaf, flower, fruit, seeds, wood and bark against these pathogens. Group of bio-compounds were determined by . red out of the ched by original copyright.

Othis item is protected by original copyright. standard phytochemical test and the antifungal bioassay were carried out on laboratory scale.

CHAPTER 2

LITERATURE REVIEW

2.1 Introduction

The residual effects of synthetic fungicide due to long usage in post-harvest management are causing health and environmental issues. Commercialized botanical or plant-based fungicides are still low in global market. Industrial and researchers are in search of novel compounds which will be able to denote a marketable technical advantage over their own or competitors' products. Furthermore, increase of fungal infection in post-harvest also aspire researcher to discover the new antifungal agents providing new mechanisms of action with a wide spectrum of antifungal activity. In our context, *Cerbera odollam* is studied in this research, in term of its antifungal activity, phytochemical and toxicity properties, as well as their bio-compounds. This chapter reviewed the bio-compounds of *Cerbera odollam*, the application and few methodology techniques employed in this study.

2.2 Cerbera odollam Gaertn

2.2.1 Botanical Description and Localization

Genus *Cerbera* belongs to the poisonous Apocynaceae family. This genus comprises of 10 to 15 species, e.g *Alstonia spatulata*, *Alstonia pneumatophora*, *Cerbera manghas* L., *Cerbera odollam* Gaertn, *Rauvolfia verticillata*, *Dyera costulata*, *Ochrosia*

oppositifolia, Anodendron nervosum Kerr, Carissa spinarum, and they are widely distributed in coastal area of South East Asia and the Indian Ocean (Kiew et al., 2011; Shen et al., 2007). In Peninsular Malaysia, there are only 2 species recorded, i.e. Cerbera odollam and Cerbera manghas (Kiew et al., 2011). This two species are often described as synonym in some literature. However, both plants are actually divided into different species by taxonomists (Abe & Yamauchi, 1977; Shen et al., 2007). They can be differentiated through the yellow-eye white corolla of Cerbera odollam and red-eye white corolla of Cerbera manghas (Cheenpracha et al., 2004).



Figure 2. 1: (A) flowers (B) fruits (C) tree of Cerbera odollam.

Cerbera odollam, commonly referred as "Pong-Pong tree" (English name) and "Buta-buta" (Malay name) is a tree that grow about 6 to 15m height (Fig. 2.1) (Kiew et al., 2011). The leaves are dark green in color. The flowers' are white and have scent reminiscent of jasmine and fruits are spherical in shape (Ahmed et al., 2008; Kiew et al.,

2011). Locations of *Cerbera odollam* in Malaysia are mostly distributed at coastal mangrove swamps, sometimes inland as a roadside tree at Perlis, Kedah, Perak, Selangor, Melaka, Pahang and Johor and are able to flower and fruit throughout the year (Kiew *et al.*, 2011).

2.2.2 Bio-compounds in Cerbera odollam

The review of bio-compounds focused on earlier and current researches. *Cerbera odollam* are well known to be rich in variety of compounds such as lignans, cardiac glycoside and terpenoids (Abe, Yamauchi, & Wan, 1989; Shen *et al.*, 2007; Yamauchi, Abe, & Wan, 1987a). Compounds structures are as shown in Fig. 2.2.

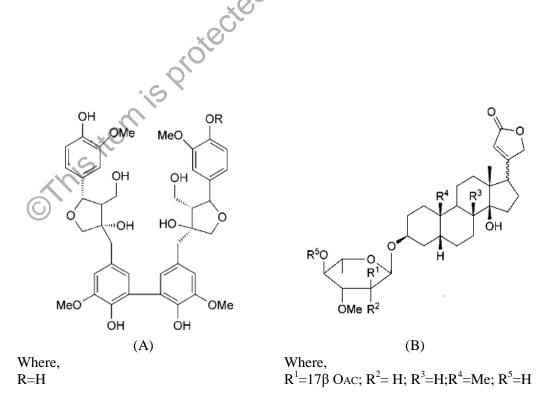


Figure 2. 2: Compounds structure of: A, Cerberalignan (lignan); B, Cerberin (cardiac glycoside) (Shen *et al.*, 2007)

The seeds are the earliest part to be investigated. Ghanekar & Ayyar (1927) had successfully detected glycorides of palmitic, staric, myristic, lignoceric, oleic and linolic acids in seed extract, followed by De Veij who had report the presence of cerberin in the seeds of *Cerbera odollam* (Bisset, 1961). The seeds are the earliest part to be investigated. The work was then further confirmed by Steldt & Chen (1943). Study of seeds was persistence until Abe & Yamauchi (1977) commenced to investigate different parts of *Cerbera odollam*, i.e. bark, leaf, stem and lignin (Abe, Yamauchi, & Wan, 1988; Abe *et al.*, 1989; Abe & Yamauchi, 1977; Yamauchi *et al.*, 1987a; Yamauchi, Abe, & Wan, 1987b). The summary of the bio-compounds identified from *Cerbera odollam* is shown in Table 2.1.

Up to date, little reports are available on the bio-compounds obtained from flower, fruits and roots. Some researchers had engaged crude extracts from flowers and fruits to test on their termite decay resistance properties on particleboard (Hashim *et al.*, 2009). Meanwhile, crude extract from roots were tested for their possibility of antinociceptive and antibacterial properties in mice (Ashikur Rahman *et al.*, 2011) and the result reported were positive. Fruits crude extract, on the other hand, failed to exhibit significant antimicrobial effects on skin bacteria (Shankar *et al.*, 2009). The current research is investigating bio-compounds from leaf, seed, stem, bark and latex. The findings on flower, fruit and roots are still limited.

Table 2.1: Summary of bio-compounds from different part of *Cerbera odollam* (leaf, seed, stem, bark and latex).

Reference Leaf Neriiforlin deacetyltanghinin 17α- deacetyltanghinin 17α- deacetyltanghinin 17α- deacetyltanghinin 17α- deacetyltanghinin 17α- deacetyltanghinin 17α- cerdollaside 17α- cerdollaside solanoside 17α- solanoside 17α- solanoside 17α- solanoside Oleagenin α-L-thevetoside (cerleaside A) Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α- neriifolin 17β- and 17α- neriifolin 17β- and 17α- deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β- and 17α- digitoxigenin Cerberidol Cerberidol 3-O-β-D-allopyranoside Cyclocerberidol Cerberidol-3-O-β-D-allopyranoside Cardiac glycoside (Laphookhieo, 2002; Laphookhieo et al., 2004; Shen et al., 2007b) Seed cerberin 2'-O-acetyl cerleaside A Cerleaside A 17β- and 17α- neriifolin Thevetin B tanghinigenin (Abe et al., 1987b) Stem Gentiobiosyl-thevetoside Tanghinigenin (Abe et al., 1987b)
deacetyltanghinin 17α- deacetyltanghinin Cerleaside A cerdollaside 17α- cerdollaside solanoside 17α- solanoside Oleagenin α-L-thevetoside (cerleaside A) Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α-neriifolin 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin 2'-O-acetyl cerleaside A 17β- and 17α- neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin Cet al., 1987b) Pongpijid, Bunyapraphatsara, & Panvisavas, 2011; Shen et al., 2007; Yamauchi et al., 1987b)
Bunyapraphatsara, & Panvisavas, 2011; Shen et al., 2007; Yamauchi et al., 1987a, 1987b) Bunyapraphatsara, & Panvisavas, 2011; Shen et al., 2007; Yamauchi et al., 1987a, 1987b) Bunyapraphatsara, & Panvisavas, 2011; Shen et al., 2007; Yamauchi et al., 1987a, 1987b) Bunyapraphatsara, & Panvisavas, 2011; Shen et al., 2007; Yamauchi et al., 1987b)
Cerleaside A cerdollaside 17α- cerdollaside solanoside 17α- solanoside Oleagenin α-L-thevetoside (cerleaside A) Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α-neriifolin 17β- and 17α-deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cyclocerberidol Cardiac glycoside Seed Seed Seed Cerleaside A 17β- and 17α- neriifolin 17β- and 17α- neriifolin 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cyclocerberidol-3-O-β-D-allopyranoside Cyclocerberidol Cardiac glycoside Seed Seed Cerleaside A 17β- and 17α- neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin (Abe et al., 1988; Shen et al., 2007; Yamauchi et al., 1987b)
cerdollaside 17α - cerdollaside solanoside 17α - solanoside 17α - solanoside Oleagenin α -L-thevetoside (cerleaside A) Oleagenin- β -glucosyl- $(1\rightarrow 4)$ - α -L-thevetoside (cerleaside B) 17β - and 17α -cerdollaside 17β - and 17α -solanoside 17β - and 17α -neriifolin 17β - and 17α -deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β - and 17α - digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol-3- O - β -D-allopyranoside Cyclocerberidol-3- O - β -D-allopyranoside Cardiac glycoside Seed cerberin $2'$ - O -acetyl cerleaside A 17β - and 17α - neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin 17β - and 17α - neriifolin Thevetin B tanghinigenin 17β - and 17α - neriifolin Thevetin B tanghinigenin 17β - 17
solanoside 17α - solanoside Oleagenin α -L-thevetoside (cerleaside A) Oleagenin- β -glucosyl- $(1\rightarrow 4)$ - α -L-thevetoside (cerleaside B) 17β - and 17α -cerdollaside 17β - and 17α -solanoside 17β - and 17α - neriifolin 17β - and 17α - deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β - and 17α - digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol-3- O - β -D-allopyranoside Cyclocerberidol-3- O - β -D-allopyranoside Cardiac glycoside Seed Seed Seed Cerleaside A 17β - and 17α - neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin et al., 2007; Yamauchi et al., 1987b) tanghinigenin
solanoside $17α- solanoside$ Oleagenin α-L-thevetoside (cerleaside A) Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) $17β- and 17α- cerdollaside$ $17β- and 17α- neriifolin$ $17β- and 17α- neriifolin$ $17β- and 17α- deacetyltanghinin$ Glucose-3-ulosyl-thevetoside $17β- and 17α- digitoxigenin$ Cerberidol Cyclocerberidol Cyclocerberidol Cyclocerberidol-3- O -β-D-allopyranoside Cardiac glycoside Seed Seed cerberin $2'-O$ -acetyl cerleaside A Cerleaside A $17β- and 17α- neriifolin$ Thevetin B Stem Gentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin et al., 1988; Shen et al., 2007; Yamauchi et al., 1987b)
Oleagenin α-L-thevetoside (cerleaside A) Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α-solanoside 17β- and 17α- neriifolin 17β- and 17α-deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin 2'-O-acetyl cerleaside A Cerleaside A 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin et al., 1988; Shen et al., 2007; Yamauchi et al., 1987b)
Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α-neriifolin 17β- and 17α-neriifolin 17β- and 17α-deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed Seed cerberin cerberin cerleaside A cerleaside A 17β- and 17α- neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin cerberin (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B tanghinigenin tanghinigenin tanghinigenin tanghinigenin
Oleagenin-β-glucosyl-(1→4)-α-L-thevetoside (cerleaside B) 17β- and 17α-cerdollaside 17β- and 17α-neriifolin 17β- and 17α-neriifolin 17β- and 17α-deacetyltanghinin Glucose-3-ulosyl-thevetoside 17β- and 17α- digitoxigenin Cerberidol Cyclocerberidol Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed Seed cerberin cerberin cerleaside A cerleaside A 17β- and 17α- neriifolin Thevetin B Gentiobiosyl-thevetoside Tanghinigenin cerberin chaptookhieo, 2002; Laphookhieo et al., 2004; Shen et al., 2007b) (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B tanghinigenin tet al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen Tanghinigenin et al., 2007; Yamauchi Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cyclocerberidol-3-O-β-D-allopyranoside Cardiac glycoside Seed cerberin (Laphookhieo, 2002; 2'-O-acetyl cerleaside A Laphookhieo et al., 2004; Cerleaside A Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen et al., 2007; Yamauchi Thevetin B Thevetin B et al., 1987b) tanghinigenin
Cardiac glycosideSeedcerberin 2'-O-acetyl cerleaside A Cerleaside A 17β- and 17α- neriifolin Thevetin B(Laphookhieo, et al., 2004; Shen et al., 2007b)StemGentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin(Abe et al., 1988; Shen et al., 2007; Yamauchi et al., 1987b) tanghinigenin
Seedcerberin 2'-O-acetyl cerleaside A Cerleaside A 17β- and 17α- neriifolin Thevetin B(Laphookhieo, et al., 2004; Shen et al., 2007b)StemGentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin(Abe et al., 1988; Shen et al., 2007; Yamauchi et al., 1987b)
2'-O-acetyl cerleaside A Cerleaside A Shen et al., 2004; Shen et al., 2007b) 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin et al., 2007; Yamauchi et al., 1987b) tanghinigenin
Cerleaside A 17β- and 17α- neriifolin Thevetin B Stem Gentiobiosyl-thevetoside Tanghinigenin Thevetin B tanghinigenin
Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen Tanghinigenin et al., 2007; Yamauchi Thevetin B et al., 1987b) tanghinigenin
Thevetin B Stem Gentiobiosyl-thevetoside (Abe et al., 1988; Shen Tanghinigenin et al., 2007; Yamauchi Thevetin B et al., 1987b) tanghinigenin
Tanghinigenin et al., 2007; Yamauchi Thevetin B et al., 1987b) tanghinigenin
Thevetin B et al., 1987b) tanghinigenin
tanghinigenin
Glocosyl-thevetosides of digitoxigenin
17α- digitoxigenin
Cerleaside B
(-)-olivil
(+)-cycloolivil
(-)-cycloolivil
Cerberalignan A
Cerberalignan B
Cerberalignan C
Cerberalignan D
Cerberalignan E
Cerberalignan F
Cerberalignan G
Cerberalignan H
Cerberalignan I