ENZYMATIC ASSISTED CITRONELLA ESSENTIAL OIL EXTRACTION FROM Cymbopogan winterianus

NORHIDATE AH BINTI ABD AZIZ

UNIVERSITI MALAYSIA PERLIS

2015



Enzymatic Assisted Citronella Essential Oil Extraction from Cymbopogan winterianus

rotected by This ten Norhidayah Binti Abd Aziz (1231110753)

A thesis submitted in fulfillment of the requirements for the degree of Master of Science in Bioprocess Engineering

School of Bioprocess Engineering UNIVERSITY MALAYSIA PERLIS

ACKNOWLEDGEMENT

First of all, Alhamdulillah, I have no words to praised the Almighty Allah, the Beneficient, the Merciful, whose blessings and exaltation flourish my thoughts to enable me to complete my master thesis. Special praises for the beloved Prophet Muhammad (P.B.U.H), who is a bonfire of knowledge and guidance for humanity.

Assoc Prof Dr. Dachyar Arbain.

My sincerest and immense gratitude to this special man, my main supervisor, for his ultimate patient, guidance, unwavering support and encouragement. Thank you for being the most understanding supervisor, the best-est of one would ever wish for. Honestly, to me he has been and will always be my great mentor; who knows multitude area of research, who always find ways to troubleshoot problems, who never failed to scare us during every meetings and discussions with his killing statement "you have to do this" "we have to add that" and "congratulations, results looks awful, need to repeat that ". Honestly, without those, I can never improved. Prof, thank you for everything. You will always be in my heart ⁽²⁾

Prof Mohd Noor bin Ahmad,

My co-supervisor. My progress tracker. Never failed to motivate me to finish early with his dedicated questions of "what's new for today" "how's progress?" but my utmost favourite is "do you need to buy anything for your research?". Prof, thank you so much for your kind support, your fatherly advice is indeed very much appreciated ©

Furthermore, I want to extend my hearted thanks to High Education Ministry of Malaysia and UniMAP for sponsoring me under Skim Latihan Akademik Bumiputra (SLAB) during this master study. The financial support, including study fees and personal allowance are highly appreciated.

Last but not least, I would like to express my utmost gratitude to my dearest husband, my backbone, Asrarul Fikri who has seen the worst to bring out the best in me. Without his precious support, i will not be able to finish my master study. Special thanks also dedicated to my beloved parents, members of the family, and to my three beautiful roses, Alisya Irdina, Hanna Qistina and our newly added family member, Rose Medina. Without their love, encouragement, support, and prayers, I could not have done anything.

Ultimately, I would like to thank all my postgraduate friends for all the support and cheers, especially to Nur Zatul 'Iffah, Nurhazwani, Azalina, Noorhidayah, Kak teh, Mubaraq, Kak Shiera, Anas, Dijah and Nabil. Without their presence, the journey of finishing master would be very challenging.

TABLE OF CONTENTS

DD			
DEC	CLARATION SHEET	i	
LIS	T OF TABLES	ii	
LIST	T OF FIGURES	iii	
LIS	T OF SYMBOLS	iv	
LIS	T OF SYMBOLS T OF ABBREVIATIONS STRAK STRACT Overview Problem Statements Research Objectives	V	
ABS	STRAK	vi	
ABS	ABSTRACT		
CHA	APTER 1 INTRODUCTION	1	
1.1	Overview	1	
1.2	Problem Statements	5	
1.3	Research Objectives	6	
1.4	Hypothesis	7	
1.5	Scope of Research	7	

CHAPTER 2 LITERATURE REVIEW

2.1	Citronella Oil	10
2.2	Global Production of Citronella Oil	11
2.3	Citronella Oil Industry in Malaysia	11

2.4	Potent	ial Use of Citronella Oil	13
	2.4.1	Insect Repellent and Pest Control Products	13
	2.4.2	Perfumery, Pharmaceutical and Aromatherapy Industries	14
2.5	Cultiv	ation of Citronella Grass	15
2.6	Harve	esting and Citronella Oil Extraction	15
2.7	The C	omposition of Lignocellulosic Biomass	17
	2.4.2	Cellulose Hemicellulose Lignin ne Assisted Extraction of Essential Oil	18
	2.4.3	Hemicellulose	19
	2.4.4	Lignin	20
2.8	Enzyn	ne Assisted Extraction of Essential Oil	22
	2.5.2	Physical Pre-Treatment	22
	2.5.3	Enzymatic Pre-Treatment	23
	2.5.4	Microbial Cellulases	25
2.9	Desig	n of Experiment (DoE)	29
	2.9.1	Response Surface Methodology (RSM)	29
	2.9.2	Central Composite Design (CCD)	31
	\bigcirc		
CHA	APTER 3	3 RESEARCH METHODOLOGY	33
3.1	Sampl	e Materials	34
3.2	Resear	rch Flow	35
3.3	Chara	cterization of Lignocellulosic Contents of Dried Citronella Roots	38
		3.3.2.1 Extractives	38

		3.3.2.2 Hemicellulose	39
		3.3.2.3 Lignin	40
		3.3.2.5 Cellulose	40
3.4	Screen	ing and Isolation of Cellulase Producing Microbes	41
	3.4.1	Soil Sample and Microorganism	41
	3.4.2	Screening and Identification of Cellulase Producer	42
	3.4.3	Growth Conditions	42
3.5	Enzyn	ne Pre-Treatment and Laboratory Scale Oil Extraction	43
3.6	Analys	sis of Crude Cellulase Activity	44
	3.6.1	Glucose Standard Calibration Curve	44
	3.6.2	Cellulase Assay	45
	3.6.3	Calculation of Cellulase Activity	47
3.7	Analy	sis of Surface Morphology using SEM	47
3.8	Pheno	typic Characteristic and Molecular Identification	48
	3.8.1	Phenotypic and Microscopic Examination of Fungi	48
	3.8.2	Molecular Identificaiton of Fungi	48
(0	3.8.2.1 Phylogenetic Tree Analysis	49
3.9	Optim	isation of Cellulase Production by Aspergillus nomius H5 strain	50
	3.9.1	Response Surface Methodology	50
	3.9.2	Optimisation of Enzyme Production	53
3.10	Optim	ization of Enzymatic Pre-Treatment and Oil Extraction	54
	3.7.1	Response Surface Methodology	54

	3.7.2	Optimization of Enzymatic Pre-Treatment and Oil Extraction	56
3.11	Gas Ch	romatography Analysis of Citronella Oil	57

CHAPTER 4 RESULTS AND DISCUSSIONS 58 4.1 Characterization of Dried Citronella Root 58 4.1.1 Lignocellulosic Contents 58 Screening and Isolation of Cellulase Producing Microorganism 4.1.2 61 idinal col 4.1.3 Construction of Growth Curve 64 4.1.4 Cellulase Activity Assay 67 Laboratory Scale Oil Extraction 4.1.5 70 Identification of Cellulase Producing Microbe 4.2 4.2.1 Macroscopic and Microscopic Identification 71 Genomic Identification 4.2.2 73 SEM Morphology Analysis of Pre-Treated Vetiver Root 4.3 76 Optimization of Cellulase Production by Aspergillus nomius H5 strain 79 4.4 79 4.4.1Statistical Analysis 4.4.2 Confirmation of Experiments and Adequacy of the Models 84 4.4.3 Effects of Parameters 86 4.4.3.1 87 pН 89 4.4.3.2 Temperature 90 4.4.3.3 **Agitation Rate**

4.4.4Interaction between Parameters91

	4.4.5	Summary of the Results	97
	4.4.6	Optimal Design	97
4.5	Optim	ization of Enzymatic Pre-Treatment and Citronella Oil Extraction	99
	4.5.1	Statistical Analysis	99
	4.5.2	Confirmation of Experiments and Adequacy of the Models	104
	4.5.3	Effects of Parameters	106
		 4.5.3.1 Enzyme Ratio (%v/v) 4.5.3.2 Reaction Time 4.5.3.3 Extraction Time Interaction between Parameters 	107
		4.5.3.2 Reaction Time	108
		4.5.3.3 Extraction Time	109
	4.5.4	Interaction between Parameters	110
	4.5.5	Summary of the Results	114
	4.5.6	Optimal Design	114
4.6	Gas Cl	hromatography Analysis of Citronella Oil	116
		is t	
СНА	PTER 5	5 CONCLUSIONS & FUTURE RECOMMENDATIONS	
5.1	Conclu	isions	116
5.2	Future	recommendations	118
REF	REFERENCES 121		

APPENDIX A Chemicals, materials, and equipments used in the study 135

APPENDIX B	Preparation of DNS solution and 50 mM sodium citrate buffer, pH 4.8	137
APPENDIX C	Analysis of cellullase activity	138
APPENDIX D	Gas Chromatography Analysis of Citronella Oil	140
APPENDIX E	Finding Summary	141
LIST OF AWAR	RDS AND PUBLICATIONS	142

orthis term is protected by original copyright

LIST OF TABLES

NO.		PAGE
2.1	Previous works on enzyme pre-treatment for extraction of various plant bioactives	23
3.1	Glucose dilution series from 10 mg/ml glucose stock solution	45
3.2	Range of variables for the CCD design for optimisation of cellulase production	51
3.3	The CCD experimental design for optimisation of enzyme production in coded and actual values	52
3.4	The goal and limits of the factors and response of the optimized cellulase production	53
3.5	Range of variables for the CCD design for enzymatic pre-treatment and oil extraction	54
3.6	The CCD experimental design for enzymatic pre-treatment and oil extraction in terms of coded and actual values	55
3.7	The goal and limits of the factors and response of the optimised pre- treatment process to enhance the citronella oil recovery	56
4.1	Composition of lignocellulosic components of dried citronella roots in percentage (dry weight)	61
4.2	Design and response of the CCD for cellulase activity level (FPase U/ml) obtained from the optimisation of enzyme production.	83
4.3	ANOVA for Response Surface Quadratic Model Analysis of variance table for optimisation of cellulase production	84
4.4	Statistical parameters obtained from ANOVA of optimise parameters for cellulase production	85
4.5	Experimental design results for pre-treatment and oil extraction of pre- treated citronella roots	101
4.6	ANOVA for quadratic model for enzymatic pre-treatment and oil extraction of pre-treated citronella roots	102
4.7	Statistical parameters obtained from ANOVA of optimisation of pre- treatment and oil extraction of pre-treated citronella roots	102

LIST OF FIGURES

NO.		PAGE
2.1	Location and arrangement of cellulose microfibril, hemicelluloses and lignin in plant cell wall (Murphy & Carthy, 2005)	17
2.2	Distribution of lignin, hemicellulose and cellulose in the secondary wall. S1, S2, S3 represent the layer of cell wall. P (primary wall) ML (middle lamella) (Kirk & Kullen, 1998).	18
2.3	Chemical structure of cellulose (Zhen Fang, 2013)	19
2.4	Chemical structure lignin (Sara Helmberger 2009)	19
2.5	Chemical structure lignin (Sara Helmberger, 2009)	20
2.6	Monomer units of lignin (Wang et al., 1992 & Chabannes et al., 2001)	21
2.7	Lignocellulosic structure before and after pretreatment (Hsu et al., 1980)	21
2.8	Structure of cellulose polymer. Crystalline cellulose discriminated from amorphous cellulose (black) by colours. Principle cellulase sites of action on the cellulose polymer liberating glucose are presented (Veeresh & Jin, 2014)	26
2.9	Layout of the Central Composite Design (CCD) of 3 variables at 5 levels (Myers, 1979)	32
3.1	Citronella plant, Citronella root and dried Citronella root used in the study.	34
3.2	Flowchart of the preliminary studies involving the isolation, and characterization of microbes of interest	36
3.3	Flowchart of the 2 nd main study involving optimisations of crude enzyme production and oil extraction.	37
4.1	Zone of clearance of the four colonies grown on the same plate.	64
4.2	Cellulolytic Indexes (CI) of the isolates.	64
4.3	Mycelial dry weight (g/100ml) and glucose consumption profiles of strains (a)UniMAPF7 (b)UniMAPF16 (c) UniMAPF24 (d)UniMAPF27	66

4.4	Mycelial dry weight (g/100ml) and cellulase activity profiles (FPase U/ml) of strains (a)UniMAPF7 (b)UniMAPF16 (c) UniMAPF24 (d)UniMAPF27	69
4.5	Laboratory scale extraction of control and pre-treated citronella root	72
4.6	Macroscopic and microscopic presentation of strain UniMAPF7 grown on PDA agar, at 37°C for 4 days.	74
4.7	Neighbour-joining phylogenetic tree of fungal sample UniMAPF7 isolate relative to members of related genus	76
4.8	Scanning Electron Microscopy images of control and pre-treated citronella roots.	80
4.9	Normal probability of internally studentized residuals of parameters for cellulase production (0.00140.2551 colour points by value of cellulase activity)	86
4.10	Plot of internally studentized residuals vs predicted response for parameters for cellulase production (0.0014 0.2551 colour points by value of level of cellulase activity)	87
4.11	One factor plot for (a) pH (b) temperature, and (c) agitation rate optimisation of paramaters for cellulase production	88
4.12	 3-D contour plot showing the effects/ interactions of pH and temperature on cellulase activity (Design point above predicted value ●; design point below predicted value ●) 	94
4.13	3-D contour plot showing the effects/ interactions of pH and agitation rate on cellulase activity (Design point above predicted value ●; design point below predicted value ●)	95
4.14	3-D response surface plot showing the effects/ interactions of temperature and agitation rate on cellulase activity (Design point above predicted value ●; design point below predicted value ●)	96
4.15	Ramps of pH, temperature, agitation rate, and cellulase activity (Desirability=1.000)	98
4.16	Normal probability of internally studentized residuals for enzymatic pre-treatment and oil extraction of pre-treated citronella root (1.2 4.8 colour points by value of citronella oil yield)	104
4.17	Plot of internally studentized residuals vs predicted response for pre- treatment and oil extraction of pre-treated citronella root (1.2	105

4.18	One factor plot for (a) ratio $(\% v/v)$ (b) reaction time and (c) extraction time in enzymatic pre-treatment and extraction of pre-treated citronella root.	106
4.19	3-D contour plot showing effects/ interactions of enzyme ratio and reaction time on oil yield of enzyme assisted extraction of pre-treated citronella root (Design point above predicted value ●; design point below predicted value ●)	111
4.20	3-D response surface plot showing effects/ interactions of extraction time and enzyme ratio on oil yield of enzyme assisted ectraction of pre-treated citronella root (Design point above predicted value ●; design point below predicted value ●)	112
4.21	3-D response surface plot showing effects/ interactions of extraction time and reaction time on oil yield of enzyme assisted extraction of pre-treated citronella root. (Design point above predicted value ●; design point below predicted value ●)	113
4.22	Ramps of ratio, reaction time, extraction time, and oil yield (Desirability=0.983)	115
4.23	Gas Chromatography (GC) analysis of pre-treated citronella oil	116
(\otimes	

LIST OF SYMBOLS

- Alpha (axial distance from centre point which makes the А design rotatable)
- Regression coefficients for the intercept coefficient β_0
- Regression coefficients for the linear coefficient βi
- Regression coefficients for the quadratic coefficient β_{ii}
- ents original convitoni Regression coefficients for the interaction coefficient β_{ij}
- °C Celsius
- Σ Standard deviation
- Coded independent variables χ_i,
- χ_j Ε Residual associated to the experiments
- G Gram
- hrs Hour
- K Number of variable
- Μ Molar
- mili Molar mМ
- μm micro meter
- Minute min
- Number of measurements Ν
- Rotation per minute rpm
- V Volume
- Volume to volume ratio v/v
- Volume to weight ratio v/w

LIST OF ABBREVIATIONS

ANOVA	Analysis of variance
Avg.	Average
BG	ß-glucosidase
СВН	Cellobiohydrolases
CCD	Central Composite Design Carbon dioxide concentration Coefficient variation Deoxyribonucleic acid
CO ₂	Carbon dioxide
conc.	concentration
CV	Coefficient variation
DNA	Deoxyribonucleic acid
DoE	Design of Experiment
FP	Filter paper
FPU	Filter paper unit
EG	Endoglucanases
ET	Extraction time
GM .S	Growth medium
H+	Hydrogen ion
HMF	Hydroxymethyl furfural
H ₂ O	Dihydrogen oxide (water)
H_2SO_4	Sulfuric acid
ITS	Internal transcribed spacer
KBr	Potassium bromide
КОН	Kalium hydroxide

- Nutrient agar NA
- not available n.a
- Hydroxide ion OH^{-}
- Oil palm fruit bunch OPFB
- PCR Polymerase chain reaction
- Potato dextrose agar PDA
- pe copyright or internis protected by original copyright RSM
- RT
- SSB
- SEM
- U
- Vs

ABSTRAK

Ekstraksi Pati Minyak Serai Wangi dari *Cymbopogan winterianus* dengan Bantuan Enzim

Minyak pati serai wangi kebiasaannya dihasilkan melalui proses penyulingan air atau stim dari akar serai wangi. Namun begitu, isipadu minyak yang dihasilkan secara umumnya agak rendah, iaitu di sekitar 0.22% - 1.0%. Keadaan ini membebankan para pengusaha minyak serai wangi untuk memenuhi permintaan tinggi terhadap penggunaan minyak tersebut. Oleh sebab itu, kajian ini dijalankan untuk meningkatkan kadar penghasilkan minyak dengan memperkenalkan penggunaan enzim mikrob yang didapati di sekitar akar serai wangi untuk diaplikasi dalam proses rawatan enzim sebelum proses pengekstrakan pati minyak. Pertama sekali, komposisi lignoselulosa akar serai wangi diselidiki bagi mengenalpasti jenis enzim yang sesuai untuk digunakan dalam proses rawatan awal akar serai wangi. Analisis komposisi kimia telah mengenalpasti komponen selulosa sebagai komponen terbanyak (38.21%±1.65), diikuti oleh hemiselulosa (30.49%±0.93), lignin (21.12%±1.86) and ekstraktif (4.97%±0.22). Sejumlah 31 daripada 52 spesis mikrob yang diasingkan telah dikenalpasti positif dalam penghasilan enzim selulase. Berdasarkan kepada indeks selulolitik vang memberangsangkan, Strain UniMAPF7, UniMAPF16, UniMAPF24, dan UniMAPF27 telah dipilih untuk penghasilan enzim mentah bagi ekstraksi pati minyak serai wangi pada skala makmal. Keputusan dari kajian analisis pertumbuhan mikrob dan kadar aktiviti enzim telah mencadangkan masa optima untuk proses penuaian enzim ialah pada hari ke 5 bagi UniMAPF7, hari ke 4 bagi UniMAPF24 dan hari ketiga bagi UniMAPF16 and UniMAPF27. Ekstraksi minyak dari akar yang telah dirawat dengan enzim semulajadi dari UniMAPF7 and UniMAPF24 menghasilkan peningkatan paling ketara masing-masing dengan pertambahan sebanyak 2.5 dan 1.7 kali ganda. Imej imbasan elektron mikroskop (SEM) bagi analisis morfologi akar yang telah dirawat membuktikan peningkatan keamatan liang-liang pori dan kerapuhan struktur fiber yang amat ketara. Analisa makroskop, mikroskop dan genomik telah mengenalpasti strain UniMAPF7Aspergillus nomius H5 strain (Nombor akses GenBank JF16646). Berdasarkan kepada parameter yang telah dikenalpasti, kaedah respons permukaan (RSM) berdasarkan reka bentuk komposit berpusat (CCD) telah diguna pakai untuk mengoptimumkan kondisi bagi penghasilan enzim selulase. Keadaan optimum untuk penghasil enzim sellulase adalah pada pH 4.73, suhu 31.7°C, dan kadar agitasi sebanyak 147rpm dengan aktiviti enzim maksimum sebanyak 0.2559U/ml. Hidrolisat enzim selulase yang optima kemudiannya digunakan untuk mengoptimumkan keadaan proses pra-rawatan enzim dan ekstraksi minyak pati. Di bawah keadaan optimum (perkadaran enzim, masa tindakbalas, masa ekstraksi), hasil minyak pati maksimum sebanyak 71.21 g/100 g substrat kering telah dicapai. Analisis varian (ANOVA) menunjukkan bahawa model dan semua parameter dianggap penting secara statistik pada 95% untuk keduadua kajian pengoptimuman menggunakan persamaan polinomial peringkat kedua. Selain itu, pengesahan model menunjukkan perkaitan yang rapat antara keputusan eksperimen dan ramalan respon. Oleh itu, model-model ini boleh digunakan dengan jayanya untuk mengenal pasti kombinasi yang berkesan daripada tiga faktor yang berbeza di dalam kedua-dua kajian pengoptimuman untuk meramalkan hasilan minyak pati dari akar serai wangi yang dirawat dengan enzim mikrob.

ABSTRACT

Enzymatic Assisted Citronella Essential Oil Extraction from Cymbopogan winterianus

The essential oil of Citronella is commonly extracted via typical steam distillation of citronella plan parts. However, the oil yield is generally low (0.22% - 1.0%), causing burden to many entrepreneurs to meet the strong demand. Therefore, the study was conducted to improve the yield of citronella oil recovery by introducing an optimised enzymatic assisted pre-treatment using crude microbial enzymes isolated from the soil surrounding the citronella roots. Firstly, the lignocellulosic components of dried citronella roots were investigated to evaluate the most abundance component for selection of enzymes useful for pre-treatment. The composition analysis has identified cellulose as the major lignocellulosic component $(38.21\% \pm 1.65)$, followed by hemicellulose (30.49%±0.93), lignin (21.12%±1.86) and extractives (4.97%±0.22). 31 out of 52 isolated strains have been identified as positive cellulase producer. Based on the calculated cellulolytic indexes, 4 strains i.e; UniMAPF7, UniMAPF16, UniMAPF24, and UniMAPF27 were selected for application in the laboratory scale pretreatment process. Results from the growth curve analysis and enzyme activity study suggested that the optimal time for enzyme harvesting occurred at day 5 for UniMAPF7, day 4 for UniMAPF24 and day 3 for UniMAPF16 and UniMAPF27. Laboratory scale oil extraction of pre-treated citronella root with crude enzymes of UniMAPF7 and UniMAPF24 yielded in a significant 2.5-fold and 1.7-fold increased in citronella oil recovery, respectively, when compared to the untreated citronella root as control. The scanning electron microscope images (SEM) of the corresponding pretreated roots with crude UniMAPF7 cellulases illustrates an increased in pore intensity and the loosening structures of root fibers are clearly evident. Macroscopic, microscopic and genomic analysis of UniMAPF7 has identified the strain as Aspergillus nomius H5 strain (GenBank accession JF16646). With known parameters range, the Response surface methodology (RSM) based on Central Composite Design (CCD) was adopted to optimize the conditions for production of crude cellulase. The optimum conditions were found to be of pH 4.73, temperature of 31.7°C, and agitation rate of 147rpm with maximum FPase activity of 0.2559U/ml. The optimised crude cellulases were further used in the optimisation of pre-treatment conditions and oil extraction process. Under optimized conditions (enzyme ratio 15.88% v/v, reaction time of 8,57hrs, and extraction time of 6.51 hrs), a maximum oil yield of 4.7397 ml/100g dry roots was achieved. The Analysis of Variance (ANOVA) test revealed that the model and all independent parameters were considered statistically significant at 95% for both optimization studies using the second order polynomial equation. The model validation showed a good agreement between experimental results and the predicted responses. Therefore the models could be successfully used to identify the effective combinations of the three different factors in both optimization studies for predicting the oil yield from pre-treated citronella roots.

CHAPTER 1

INTRODUCTION

1.1 Overview

Cymbopogan winterianus, also known as serai wangi or citronella grass is native to Sri Lanka and Sounth India (Ahmed, 2005). It can also be found growing wild in most tropical countries such as Malaysia, Thailand and Indonesia. Recently, this plant has been planted commercially by enterprenours for its essential oil, commercially known as Citronella oil (Azmil *et al.*, 2005)

Citronella oil holds an important position in essential oil industry. The oil, which can be obtained from the leaves and roots of *Cymbopogan winterianus*, is regarded as one of the twenty most important essential oils found in the world trade (Lawrence, 1993). It contains unique chemical compounds such as citronellol, citronellal and geraniol which are widely used in perfumery, aromatherapy, detergent, cleaning compounds and other industrial products (Rosalinda & Tio, 2001). Apart from that, the oil has also being recognized as a potent insecticide and is widely used as the main ingredient in many insect repellent products (Pinheiro *et al.*, 2013)

The current world demand and price for Citronella oil is experiencing fluctuation due to proliferation of inexpensive synthetic isolates derived from turpentine oil and *Eucalyptus citriodara* oil (Katiyar, 2011). These synthetic isolates were generally cheaper in price, making them more preferable whenever the only criterion of choice is the price (Tiwari, 2010). However, the quality of natural citronella oil remains

superior compared to the synthetic one (Rosalinda & Tio 2001). Such property explains the strong demand exist for natural citronella oil as it is still the preferred choice for application in perfumery industry mainly because of its unique and stable properties which are vital in blending perfumes and compounding industrially important essences (Lawrence, 2004).

Presently, world production of citronella oil is approximately 5000 tonnes, valued at 20million USD per year (Tiwari, 2010). The major producing countries are China, Indonesia, Taiwan, Malaysia, Brazil, Ceylon, India, Guatemala, Argentina, Ecuador, Madagascar, and Mexico (Tiwari, 2010). The oil is commonly obtained via hydrodistillation or steam distillation of *C. nardus* plant parts, particularly the roots. However, these conventional methods of oil extraction often yielded a relatively low percentage of essential oil (0.22% - 1.0% of oil recovery), indirectly resulting shortage of worldwide supply (Cassel & Vargas, 2006; Wany *et al.*, 2013). Due to the increasing demand of the oil, countless research efforts have been made to improve the extraction yield of citronella oil; either by introducing sophisticated extraction techniques or by incorporating additional enzymatic or chemical pre-treatment steps prior to oil extraction processes.

Recently, the enzyme assisted extractions are gaining preference because the need for ecofriendly extraction technologies. This method also affects the possibility of greener chemistry as pressure mounts on trying to identify cleaner routes for the extraction of new compounds. Enzymatic pre-treatment has shown to successfully improved the extraction yield of cloves, celery, thyme (*Thymus capitatus* L.) and Rosemary (*Rosemarinus officinalis* L.) without altering the physical and chemical

characteristics of the oil (Munish *et al.*, 2012). In some cases like ginger and garlic, enzymatic pre-treatment have shown a 50% improvement in essential oil yield (Munish *et al.*, 2012). Interestingly, the enzymatic extraction of vanilla pod not only improved the oil yield, but also resulted in the production of superior quality of vanilla oil (Madava *et al.*, 2012). Despite the enzymatic assisted essential oil extractions on the aforementioned plant species were shown to be successful, its application on citronella oil extraction from *Cymbopogan winterianus* remains to be explored.

The limiting factor for an efficient oil extraction is due to the fact that the structures of essential oil producing cells are found embedded inside the lignocellulosic materials of the citronella root. Therefore, the propose method for this study is to enhance oil release by using biological degradation which involves enzymatic hydrolysis of the glycosidic linkages of the cellulose chains following simple mechanical treatment on the lignin polymer. This method was proven a success in enhancing oil recovery of other species. According to Munish *et al.*, (2012), enzymatic pre-treatment degrades the cell wall into simpler molecules, resulting in partial decomposition of lignocellulosic structure to facilitate the oil flow prior to extraction.

However, the enzyme-assisted extraction of bioactive compounds from plants (citronella roots) is subjected to few potential commercial limitations. Firstly, the use of commercial enzymes in industrial scale may be hampered by changes in environmental conditions such as pH, temperature and percentage of dissolved oxygen levels, which will eventually reduce the effectiveness of pre-treatment and would incur more cost for large-scale optimisation (Kalia *et al.*, 2001). Secondly, regardless of the high activity of the commercial enzymes, it's effectiveness for pre-treatment of lignocellulosic materials

is less convincing due to incomplete hydrolysis of plant cell wall (Munish *et al.*, 2012). On the onther hand, the high cost associated with purchasing commercial enzymes for pre-treatment of large volume of raw materials in the scale-up processes are often not practical. An expensive pre-treatment method is highly not prefereable since enterprenours are currently seeking for the most practical and cost effective pre-treatment process in order to maximise profit.

Alternatively, enzymatic pre-treatment using whole microbe seems promising since it incurs a much lower cost of production (Kalia *et al.*, 2001). There are a few established commercial enzymes producing microbes may be useful for assisting the extraction of citronella oil. Nevertheless, the use of these strains may not be successful as the oil was shown to possess strong antimicrobial properties as previously reported by Lee and Wendy, 2013. Various publications have also documented similar findings that showed the antimicrobial properties of citronella oil in inhibiting the growth of many organisms including the eight medically important *Candida* species (Silva *et al.*, 2008) and the five strains of *Propionibacterium acnes* (Luangnarumitchai *et al.*, 2007).

Nevertheless, microbes have evolved to develop resistance mechanisms upon exposure to any inhibitory compounds (Cloete, 2003). Considering this, it is speculated that the above limitation can be overcome by isolating microbes that lives surrounding the citronella roots (Pattnaik, 1996). Those microbes have been exposed to the antimicrobial properties of citronella oil. Therefore, the resistance mechanism is presumed to already been established, resulting in an improve tolerance towards the inhibitory affect of the citronella oil.

1.2 Problem Statements

The multitude usages of citronella oil have caused tremendous demand for its supply. The oil, obtained via typical steam distillation of citronella plan parts, however, often yielded a relatively low amount of oil volatiles. Alternatively, an eco-friendly technology of enzyme pre-treatment using commercial enzyme was proven successfull in improving the extraction yield in other plant species. This however, subjected to few limitations. The high cost associated with commercial enzyme is of greatest concern and the use of established enzyme producing microbes on the other hand could be hampered by the antimicrobial properties of the citronella oil. Considering this, the present investigation was intended to utilise the use of microbes isolated surrounding the citronella roots for production of crude microbial enzymes that would later be optimised it's production and it's suitability for pre-treatment process.

. suitability for pre