

by

EMI SHAQIZA AZIZI (0730410181)

A thesis submitted in fulfill requirements for the degree of Master of Science (Materials Engineering)

> School of Material Engineering UNIVERSITI MALAYSIA PERLIS

ACKNOWLEDGEMENT

In the name of Allah, Most Gracious, Most Merciful

First and foremost, I would like to express my deepest gratitude to my parents for their support and never ending love. Thank you for giving me the absolute freedom to choose the path of life to walk. This achievement would serve to be one of the first great accomplishments of many more to come.

Next, with a deep sense of gratitude, I wish to express my sincere thanks to my supervisors, Professor Dr. Mohd Noor Ahmad and Dr. A.K.M. Shafiqul Islam, for their immense guidance, dedication and motivation throughout the project years. Their wide knowledge, valuable advice and encouragement have been of great value for me and will be remembered lifelong. It is a pleasure to have the opportunity to learn and work with both of you.

Besides, I would like to extend my warmest thanks to those whom have helped throughout my work especially Dr. Dahcyar Arbain and also to academic staffs and technicians of School Material Engineering, School of Bioprocess Engineering and Agrotechnology Development Unit for their kind assistance trough out my research years. It will be impossible to complete my research as scheduled without their kind arrangement especially the use of SEM, HPLC and other analytical equipments.

Last but not least, I am grateful to my friends especially in Cluster Sensor, Application and Technology (Pak Iqmal, Ridzuan, Kak Nurul, Wahyu, Azuddin, Chin, Saskya, and Fathi) for their encouragement, moral support and understanding. Finally, I would like to take this opportunity to thank all those whom I have not mentioned. Their direct and indirect assistance have had an impact to this thesis. The whole experience of pursuing this MSc programme has taught me to be more independent, patient and in understanding the true meaning of "scientific research".

ontris termis protected by original convited

TABLE OF CONTENTS

| Acknow | vledgement | ii |
|-----------|--|------|
| Table o | f Content | iv |
| List of 7 | Tables | ix |
| List of] | Figures | X |
| List of A | Abbreviations | xiii |
| List of S | Symbols | XV |
| Abstral | x (Bahasa Melayu) | xvi |
| Abstrac | et (English) | xvii |
| Chapte | r 1 OTTO | |
| 1.0 | Introduction | 1 |
| 1.1 | Overview | 1 |
| 1.2 | Molecularly Imprinted Polymer (MIP) as Selective Sorbent | 2 |
| 1.3 | Solid-Phase Extraction (SPE) | 3 |
| 1.4 | Curcumin as Phytochemicals | 4 |
| 1.5 | Research Approach | 5 |
| 1.6 | Specific Objectives | 6 |
| | | |
| Chapte | r 2 | |
| 2.0 | Literature Review | 7 |
| 2.1 | History of Molecular Imprinting | 7 |
| 2.2 | Principles of Molecular Imprinting | 9 |

2.3 Types of Molecular Imprinting 12

| | 2.3.1 | Covalent Method | 12 |
|--------|----------|--|----|
| | 2.3.2 | Non-covalent Method | 16 |
| 2.4 | Molecul | ar Recognition | 22 |
| | 2.4.1 | Template | 22 |
| | 2.4.2 | Functional Monomers | 24 |
| | 2.4.3 | Cross Linkers | 26 |
| | 2.4.4 | Porogen | 28 |
| | 2.4.5 | Polymerization | 30 |
| 2.5 | Molecul | ar Modelling for Functional Monomer Prediction | 32 |
| 2.6 | Polymer | Evaluation and Characterization | 33 |
| | 2.6.1 | Batch Binding Analysis and Isotherms | 33 |
| | 2.6.2 | Characterization of MIP by Gas Adsorption Technique | 43 |
| 2.7 | Molecul | arly Imprinted Solid-Phase Extraction (MISPE) | 46 |
| 2.8 | Curcumi | n derived from Turmeric (Curcuma longa) | 49 |
| | | | |
| Chapte | r 3 | | |
| 3.0 | Material | s and Method | 50 |
| 31 | Chemica | ıls | 50 |
| 3.2 | Stock an | d Working Standard Solutions Preparation | 50 |
| 3.3 | Molecul | ar Modelling | 51 |
| 3.4 | UV Spec | ctrophotometer Analysis | 52 |
| 3.5 | Preparat | ion of molecularly imprinted polymer (MIP) and non-imprinted | 53 |
| | polymer | (NIP) | |

| Remova | al of Curcumin from MIP using Soxhlet extraction | | | | |
|--|--|---|--|--|--|
| Porosity | y and Surface Area Analysis | | | | |
| Batch Binding Analysis | | | | | |
| 3.8.1 | A Study on the Selection of Functional Monomers and Porogen | 57 | | | |
| 3.8.2 | A Study on the Selection of Solvent for Batch Binding | 58 | | | |
| | Analysis | | | | |
| Isotherm | ns Model | 58 | | | |
| Optimization of Molecularly Imprinted Solid-Phase Extraction | | | | | |
| (MISPE | MISPE) | | | | |
| 3.10.1 | 10.1 Solid-Phase Extraction (SPE) Studies | | | | |
| 3.10.2 | Curcumin Extraction from Turmeric (Curcuma longa) | 60 | | | |
| 3.10.3 | Pretreatment of Curcumin Extraction using Molecularly | 61 | | | |
| | Imprinted Solid-Phase Extraction (MISPE) | | | | |
| 3.10.4 | Curcumin Detection using High Performance Liquid | 62 | | | |
| | Chromatography (HPLC) | | | | |
| | 3.10.4.1 Mobile Phase Preparation | 62 | | | |
| n1. | 3.10.4.2 High Performance Liquid Chromatography | | | | |
| | (HPLC) Determinations | 62 | | | |
| | Porosity Batch B 3.8.1 3.8.2 Isotherm Optimiz (MISPE 3.10.1 3.10.2 3.10.3 3.10.4 | Porosity and Surface Area Analysis Batch Binding Analysis 3.8.1 A Study on the Selection of Functional Monomers and Porogen 3.8.2 A Study on the Selection of Solvent for Batch Binding Analysis Isotherms Model Optimization of Molecularly Imprinted Solid-Phase Extraction (MISPE) 3.10.1 Solid-Phase Extraction (SPE) Studies 3.10.2 Curcumin Extraction from Turmeric (<i>Curcuma longa</i>) 3.10.3 Pretreatment of Curcumin Extraction using Molecularly Imprinted Solid-Phase Extraction (MISPE) 3.10.4 Curcumin Detection using High Performance Liquid Chromatography (HPLC) 3.10.4.1 Mobile Phase Preparation 3.10.4.2 High Performance Liquid Chromatography (HPLC) Determinations | | | |

Chapter 4

| 4.0 | Results and Discussion | 64 |
|-----|---|----|
| 4.1 | Molecular Modelling for Template – Functional Monomer | |
| | Interaction | 64 |

| 4.2 | Spectroscopic Evaluation via UV Spectrophotometer | | | |
|-----------|---|--|-----|--|
| | 4.2.1 | Template – Functional Monomer Interactions | 68 | |
| | 4.2.2 | Porogen Effect on Binding Prediction | 71 | |
| 4.3 | Polymer | Preparations and Template Removal | 73 | |
| 4.4 | Physical | Characterization using BET and BJH Analysis | 76 | |
| 4.5 | Batch Bi | inding Analysis | 78 | |
| | 4.5.1 | Functional Monomer Conformation | 78 | |
| | 4.5.2 | Porogen Conformation | 83 | |
| | 4.5.3 | Solvent Adsorption Effect on Polymer | 85 | |
| 4.6 | Isothern | rms Models | | |
| 4.7 | Molecul | larly Imprinted Solid-Phase Extraction Procedures (MISPE) | | |
| | 4.7.1 | Optimization of Solid-Phase Extraction (SPE) | 92 | |
| | | 4.7.1.1 pH Effect on Loading Solvent | 92 | |
| | | 4.7.1.2 Washing Solvent | 93 | |
| | . 0 | 4.7.1.3 Eluting Solvent | 95 | |
| | . 5 | 4.7.1.4 Polymer Mass | 96 | |
| \langle | 4.7.2 | Curcumin Extraction from Curcuma longa (Turmeric) and | | |
| | | Pretreated with Molecularly Imprinted Solid-Phase Extraction | | |
| | | MISPE | 97 | |
| Chapte | Chapter 5 | | | |
| 5.1 | Conclus | ions | 101 | |

| 5.2 Recommendations for future works | 102 |
|--------------------------------------|-----|
| 5.2 Recommendations for future works | 102 |

References

Appendices

| Appendix A: Standard Curve | |
|----------------------------|--|
|----------------------------|--|

Appendix B: Isotherm Models

onthis item is protected by original copyright

121

123

LIST OF TABLES

Table

| 2.1 | Types of non-covalent interactions with importance to molecular | |
|-----------|---|-----|
| | imprinting represented by schematic models and examples | 19 |
| 2.2 | Commonly use functional monomers for | |
| | non-covalent imprinting | 25 |
| 2.3 | Commonly use cross linking agent | 28 |
| 2.4 | Summarization of polymerization formats | 31 |
| 2.5 | Examples of MIP with binding isotherms models | 36 |
| 2.6 | LF-I coefficients for MIP1 and MIP2 | 42 |
| 3.1 | The conditions of curcumin added with functional monomer | 52 |
| 3.2 | Polymerization composition | 53 |
| 3.3 | Isotherm model equation | 59 |
| 3.4 | SPE test conditions | 60 |
| 4.1 | Binding energies ΔE of curcumin with MAA and AM | 66 |
| 4.2 | Physical properties of MIP using different porogen and functional | |
| \langle | monomer | 77 |
| 4.3 | IF value of P1 – P6 in MeOH, 24 h of incubation | 83 |
| 4.4 | Independent t-Test for MISPE and NISPE | 100 |

LIST OF FIGURES

Figure

| 2.1 | Graphical representation illustrating the number of original papers | |
|------|---|----|
| | published | 8 |
| 2.2 | Principle of molecular imprinting inspired by Fischer lock-and-key | |
| | metaphor | 10 |
| 2.3 | Schematic illustration of molecular imprinting | 11 |
| 2.4 | Schematic of covalent imprinted polymer | 13 |
| 2.5 | Covalent imprinting of mannopyranoside using its 4-vinylphenylboronic | |
| | acid ester | 15 |
| 2.6 | Schematic of non-covalent imprinted polymer | 16 |
| 2.7 | Non-covalent imprinting by theophylline | 21 |
| 2.8 | Batch rebinding approach | 34 |
| 2.9 | Scatchard plot for the binding nature of Resveratrol | 35 |
| 2.10 | Depiction of the typical binding isotherm measured in MIP (thick line). | |
| | The thin line shows the FI fit | 40 |
| 2.11 | The corresponding AD of FI | 40 |
| © ' | | |
| 2.12 | Experimental adsorption isotherm for MIP1 (a) and MIP2 (b) (data | |
| | points), and corresponding fitted LF-I (lines) | 42 |
| 2.13 | Gas absorbed into solid phase and adsorbed onto a solid surface | |
| | respectively | 44 |
| 2.14 | Adsorption isotherms according to IUPAC classifications | 45 |
| | | |

| 2.15 | Vacuum manifold SPE, packed with MIP | | |
|------|---|----|--|
| 2.16 | Basic SPE procedures; (1) Sample loading, (2) Washing off the | | |
| | interferences, (3) Eluting the template | 48 | |
| 2.17 | Structure of curcumin | 49 | |
| 3.1 | Thermal polymerization carried out in water bath | 54 | |
| 3.2 | Soxhlet extraction set up | 56 | |
| 4.1 | Molecular structure of curcumin. 🔲 : Functional group that | | |
| | able to interact with functional group in functional monomer | 66 | |
| 4.2 | Optimized conformations of curcumin, MAA and AM | 66 | |
| 4.3 | (a) and (c) Complex formed between curcumin and two molecules | | |
| | of MAA and AM, respectively. The presence of hydrogen bond is | | |
| | indicated by the dashed black line. (b) and (d) An alternative possible | | |
| | configuration of MAA and AM, respectively | 68 | |
| 4.4 | Complex formed between curcumin and four molecules of | | |
| | AM and MAA, respectively. The presence of hydrogen bonding | | |
| | indicated by the dash black line | 69 | |
| 4.5 | UV spectra of curcumin added with MAA | 70 | |
| 4.6 | UV spectra of curcumin added with AM | 72 | |
| 4.7 | Adding 4 mmol of MAA in different type of porogen | 73 | |
| 4.8 | The morphology of polymer prepared via bulk polymerization method | 75 | |
| 4.9 | Solvent absorbance (a): before template removal and (b) after template | | |
| | removal | 76 | |
| 4.10 | SEM image of MIP (a): before template removal, (b): after template | | |

removal

| 4.11 | Schematic reaction of imprinted polymer between curcumin and MAA | 80 |
|------------|---|-----|
| 4.12 | Standard calibration curve of curcumin in MeOH | 81 |
| 4.13 | Total amout of curcumin bound to polymer, Q in MeOH, initial | |
| | concentration: 3 µg/ml, 24 hours of incubation | 82 |
| 4.14 | IF value of P1 – P6 in different adsorption solvent | 87 |
| 4.15 | Curcumin binding isotherms for P1. The polymers were incubated in | |
| | MeOH with an increasing amount of curcumin for 24 h | 89 |
| 4.16 | Curcumin binding adsorption of MIP1 fits to LI | 90 |
| 4.17 | Curcumin binding adsorption of MIP1 fits to FI | 91 |
| 4.18 | Curcumin binding adsorption of MIP1 fits to L-FI | 92 |
| 4.19 | Curcumin bound (%) of loading solvent in MeOH with different pH | 96 |
| 4.20 | Effect on HOAc% in washing step to SPE cartridge | 97 |
| 4.21 | Effect on HOAc% in eluting solvent to SPE cartridge | 99 |
| 4.22 | Effect of recoveries (%) with different polymer weight (mg) | 100 |
| 4.23 | Chromatograms of spiked turmeric samples using MISPE and NISPE | 101 |
| 4.24 | Recoveries (%) of curcumin by MISPE and NISPE | 102 |
| \bigcirc | Y Contraction of the second | |

77

LIST OF ABBREVIATIONS

| % | - | Percent |
|-----------------------|------|--|
| ⁰ C | - | Degree celcius |
| µg/g | - | Microgramme per gramme |
| µg/ml | - | Microgramme per millilitre |
| μm | - | Micron, micrometre |
| μL | - | Microlitter |
| a | - | Isotherm constant |
| AD | - | Affinity distribution |
| AIBN | - | Azobisisobutyronitrile |
| AM | - | Acrylamide |
| AM1 | - | Semi empirical - quantum mechanics |
| В | - | Bound template concentration |
| BET | - | Brunauer, Emmett and Teller |
| BJH | - | Barrett-Joyner-Halenda |
| C _f | - | Free template concentration |
| Ci | - | Initial concentration |
| CHCl ₃ | - | Chloroform |
| cm^3/g | - | Cubic centimeter per gramme |
| d_m | - | Molecule diameter |
| D_p | - x0 | Pore diameter |
| E _{complex} | | Total energy of curcumin with functional |
| 1 | | monomer |
| E _{curcumin} | 2 | Total energy of curcumin |
| E _{monomer} | - | Total energy of functional monomer |
| EDMA | - | Ethylene glycol dimethacrylate |
| F | - | Final concentration |
| g | - | Gramme |
| GC | - | Gas chromatography |
| HOAc | - | Acetid acid |
| H ₂ O | - | Water |
| HPLC | - | High performance liquid chromatography |
| IF | - | Imprinting factor |
| K_0 | - | Median association constant |
| L | - | Litre |
| MeCN | - | Acetonitrile |
| MeOH | - | Methanol |
| FI | - | Freundlich isotherm |
| LI | - | Langmuir isotherm |
| L-FI | - | Langmuir-Freundlich isotherm |
| m | - | Heterogeneity index |
| mg | - | Milligramme |

| m^2/g | - | Square meter per gramme |
|------------------|------|--|
| MIP | - | Molecularly imprinted polymer |
| MISPE | - | Molecularly imprinted solid-phase |
| | | extraction |
| ml | - | millilitre |
| mmol | - | Millimole |
| mm Hg | - | Millimeters of mercury |
| NIP | - | Non-imprinted polymer |
| nm | - | Nanometer |
| N _t | - | Total number of binding sites |
| \mathbf{p}_0 | - | Saturation pressure of the gas |
| р | - | preassure |
| Q | - | Total binding amount |
| Q _{MIP} | - | Total binding amount of curcumin towards |
| | | MIP |
| Q _{NIP} | - | Total binding amount of curcumin towards |
| | | NIP |
| SD | - | Standard deviation |
| SEM | - | Scanning Electron Micropgraph |
| SPE | | Solid-phase extraction |
| SPME | O | Solid-phase microextracion |
| THF | - | Tetrahydrofuran |
| UV | - x0 | Ultraviolet |
| V | -0- | Volume |
| V _m | 5 | Quantity of the gas adsorbed at pressure |
| w .S | ×, | Weight |
| w/v | - | Weight/volume |
| | | |
| | | |
| | | |
| | | |
| | | |
| \bigcirc | | |

LIST OF SYMBOLS

| ΔE | - | Binding energy | |
|-----------------------|--------------------|--|-------|
| $\sum_{n=1}^{\infty}$ | - | Summation | |
| π^* | - | Excited state | |
| ۸ ٧ | - | wave length | |
| % | - | Percentage | |
| | | | . 6, |
| | | | 1 CTC |
| | | -0 | 3 |
| | | COX | / |
| | | | |
| | | | |
| | | | |
| | | 10 | |
| | | , Or | |
| | | the second secon | |
| | | | |
| | xe | × | |
| | | | |
| | XO | | |
| | XO | | |
| | $\hat{\mathbf{Q}}$ | | |
| | | | |
| | | | |
| ٠ | XOY | | |
| | \mathbf{N} | | |
| |) | | |
| (Y | | | |
| | | | |
| | | | |
| | | | |

ABSTRAK

PENCIRIAN POLIMER BERCETAK MOLEKUL UNTUK PENGEKSTRAKAN KURKUMIN DARI Curcuma longa (Kunyit)

Polimer molekul bercetak (MIP) untuk pengekstrakan selektif kurkumin telah disentesis melalui ikatan bukan kovalen di mana kurkumin digunakan sebagai templat. Polimer dibina dengan menggunakan 2 jenis monomer berfungsi iaitu asid metakrilik (MAA) dan akrilamida (AM) dengan 3 jenis pelarut iaitu klorofom (CHCl3), tetrahidrofuran (THF) dan asitonitril (MeCN). Analisis keluasan permukaan dan keporosan menunjukkan polimer yang dibina menggunakan THF memberikan saiz dimater pori tertinggi iaitu 618.43 nm. Proses pengikatan semula menunjukkan faktor molekul bercetak tertinggi adalah pada polimer yang dibina daripada MAA dan THF. Pencirian seterusnya menggunakan 3 jenis model isoterm iaitu Langmuir (LI), Freundlich (FI) dan Langmuir-Freundlich (LF-I). Solver daripada Micrsoft Excel telah digunakan untuk mengenalpasti nilai parameter dalam setiap isoterm dengan mengoptimumkan nilai R². Nilai R² yang dihitung adalah 0.91 (LI), 0.69 (FI) dan 0.96 (LF-I). Ruang bercetak, N dan kehomogenan, m telah dihitung bagi MIP dan polimer molekul tidak bercetak (NIP) menggunakan LF-I. Didapati N bagi MIP lebih tinggi iaitu 1250.62 µg/g berbanding NIP 998.35 µg/g. Analisis ini menunjukkan bahawa MIP mempunyai lebih ruang bercetak dan selektif terhadap kurkumin. Seterusnya, pengekstrakan fasa pepejal (SPE) telah dilakukan dengan menggunakan polimer seberat 150 mg. Analisis ini bertujuan untuk mengekstrak kurkumin dan hasil menunjukkan nilai permerolehan semula bagi MIP adalah 43.10 %, lebih tinggi berbanding NIP iaitu 13.46%. Bagi tujuan pengekstrakan kurkumin daripada kunyit, analisis yang sama telah dijalankan dan pemerolehan semula bagi MIP adalah sebanyak 67.76% dan NIP adalah 39.86%. Ini menunjukkan bahawa penghasilan MIP mempunyai potensi yang tinggi bagi tujuan purifikasi kurkumin dengan mengaplikasikan SPE.

ABSTRACT

CHARACTERIZATION OF MOLECULARLY IMPRINTED POLYMER FOR AN EXTRACTION OF CURCUMIN FROM *Curcuma longa* (Turmeric)

A molecularly imprinted polymer (MIP) for selective extraction of curcumin has been synthesized via non-covalent approach by using curcumin as a template. Polymerization was prepared using 2 (two) functional monomers namely methacrylic acid (MAA) and acrylamide (AM) together with 3 (three) different porogens namely chloroform (CHCl₃), tetrahydrofuran (THF) and acetonitrile (MeCN). Porosity and surface area analysis revealed that the polymer prepared using THF as porogen has the highest average pore diameter size i.e. 618.43 nm. Batch binding analysis revealed that the largest imprinting factor was attained by the polymer prepared using MAA and THF as functional monomer and porogen respectively. Further characterization was carried out using 3 (three) isotherm models namely Langmuir (LI), Freundlich (FI) and Langmuir-Freundlich (LF-I) isotherm. The unknown parameters in each isotherm were calculated by using Solver function in Microsoft Excel and were optimized for R^2 value. The calculated R^2 values were found to be (0.91), (0.69) and (0.96) for LI, FI and LFI respectively.. Hence, LFI was further used to calculate the binding sites (N) and homogeneity (m) of both the MIP and NIP (non-imprinted polymer). The result showed that MIP1 has more Nt (1250.62 $\mu g/g$) as compared to NIP (998.35 $\pi g/g$) suggesting that MIP has more binding sites and selective towards curcumin. A 150 mg of polymer mass was packed into SPE (solid phase extraction) cartridge and subsequently used to extract curcumin from raw turmeric extract. The recoveries were 43.10% for MIP as compared to 13.46% for NIP. This suggested that the MIP cartridge exhibited significant selectivity toward curcumin, with recoveries 67.76% and 39.86% for NIP, indicating that the synthesized MIP has the potential for curcumin purification through SPE.

CHAPTER 1

INTRODUCTION

1.1 Overview

In general agreement, bioactive compounds are essential and non-essential compound (e.g. vitamins or polyphenol) that derived naturally, are part of food chain and could give an effect on human health (Biesalski et al., 2009). These compounds may exert their effect by acting as antioxidants, activating liver detoxification enzymes, blocking the activity of bacterial or viral toxins, inhibiting cholesterol absorption, decreasing platelet aggregation, or destroying harmful gastrointestinal bacteria (Pennington, 2002). Nowadays, consumers are much aware in nutrition value and food fortification for healthcare. The abundance of traditional medicines and supplements proved that the market has high demands on natural products.

Because of the high demand, it should have a scientific way in handling and preparing bioactive compounds before it could be used for alternative products. The process of handling natural products is normally tedious and time consuming. Currently, many researches are conducted for final separation and detection steps, while less attention are paid to the development of faster, more selective cleaned up method (Möller, 2006). Sample cleaned up is very important for samples with complex matrices, such as biological fluids, food extracts and wastewater. This is because complex matrices usually contained various compounds that might suppress the targeted analytes signal. Before injection into a liquid chromatography system or other analytical equipments, the sample matrix must be separated from the analytes of interest. Otherwise, contaminants could disturb separation and detection or even damage the analytical column. The cleaned up procedure also could enriched the analytes concentration which will improve the sensitivity.

Cleaned up method depends on types of matrices and type of targeted compound (volatile or non-volatile). Commonly, for non-volatile compound, an extraction procedure using solid-phase extraction (SPE) could be carried out. A combination of SPE with selective sorbent is a great system which could fasten up the procedure. Selective sorbent is produced using molecularly imprinted polymer (MIP) which is build based on the targeted compound. In this study, the preparation of MIP for curcumin is developed and implemented in SPE.

1.2 Molecularly imprinted polymer (MIP) as selective sorbent

Recently MIP has been broadly used as a selective sorbent or as a stationary phase for the extraction of various drugs, natural substances or traditional medicinal compounds (Rashid, Briggs, Hay, & Stevenson, 1997; Wensheng & Gupta, 2004; Yinzhe & Kyung, 2006). This technique is an adaptation from the role of antibodies antigents concept and was developed as synthetic receptors which are suitable for separation technique. This was based on tailored selectivities and affinities. In general, an objective substrate is employed as a template molecule that binds with functional monomer during the copolymerization of the functional monomer and the cross linking agent. Movements of molecules are frozen in polymeric structures so that they are immobilized in a desired fashion (Komiyama, Takeuchi, Mukawa, & Asanuma, 2003b). Removal of the template during washing step will leave behind cavities on the polymer matrix with the shape of the selected template. The arrangement of functional groups will result in complementary binding sites to the original template. The MIP can be prepared using covalent and non-covalent method and however the latter approach is more widely used because of its simplicity (Wang, Hong, & Row, 2004). This technique has been introduced since 1970s and developing tremendously until present. Some of the advantages of this technique are: 1) the preparation is straightforward, 2) the MIP itself is stable in terms of mechanically and thermally and 3) the MIP is reusable.

1.3 Solid-phase extraction (SPE)

Solid-phase extraction (SPE) is one of the applications that use MIP as a selective sorbent. Typically in SPE, solutes are extracted from a liquid phase into the solid phase which are readily packed with porous particles of silica with a bonded organic phase or of an organic polymer such as cross linked polystyrene (Fritz, 1999). Conventional SPE has some limitation especially in terms of specificity. With the development of MIP, the empty SPE cartridge can be packed with optimized MIP. The selective cartridge is suitable for preconcentration and sample cleaned up before analyzing using chromatographic technique. This would be a good application for specific analysis. Once MIP is optimized, it will be packed in empty SPE cartridge, followed by optimization of SPE procedures which involves conditioning the cartridge, loading samples, washing the interferences and finally eluting the desired template. This technique also knows as molecularly imprinted solid-phase extraction (MISPE) which was firstly done to clean up pentamidine from urine (Börje, 1994). The MISPE procedures are similar to conventional SPE procedures (Caro, Marcé, Borrull, Cormack, & Sherrington, 2006). Thus, it has to be optimized to achieve maximum template recovery. The choices of solvent during MISPE procedure is crucial and need to be done carefully so that the binding interaction occurred during sample loading will not diminished and finally affected the selectiveness.

1.4 Curcumin as phytochemicals

Curcumin derived from turmeric is categorized as low molecular weight of polyphenol which is one the phytochemicals that obtained naturally in plants. Phytochemicals derived from dietary components has gained much attention to threat human diseases especially cardiovascular diseases and cancer. Curcumin has gained vast acknowledgements as an antioxidant in health and medicinal fields apart from the main role as the food colorant and preservatives (Aggarwal, Surh, & Shishir, 2007; Basile et al., 2009; Gopinath et al., 2004; Hailong & Qingrong, 2010; Manikandan, Sumitra, Gayathri, & Lonchin, 2006; Motterlini, Foresti, Bassi, & Green, 2000).

1.5 Research approach

The main objectives of this study are divided into two sections; the development studies of MIP and its application on SPE technique. The fundamental studies of MIP are important in order to produce the most effective MIP towards curcumin. A good MIP resulted with high imprinting factor which is lead to high selectivity towards target molecule. The study begins by selecting suitable components for polymerization which are functional monomer and porogen. An effective MIP is the outcome from good combination of template - functional monomer porogen. Once an effective MIP was selected, it was than implemented in SPE. Solid-phase extraction involves four basic procedures which are conditioning, loading, washing and eluting. To achieve maximum recovery of template during SPE, it was optimized using different conditions of loading, washing, eluting solvent and mass polymer. The molecularly imprinted solid-phase extraction (MISPE) was used to extract curcumin as the bioactive compound from turmeric. Characterizations were done using analytical equipments and supported with binding isotherms.

1.6 Specific objectives

The objectives of this study are as follows:

- To develop MIP that is selective towards curcumin
- To characterize the curcumin-imprinted polymer, particularly on functional monomer and porogen
- To apply the characterized MIP for extraction of curcumin using MISPE scheme

6

CHAPTER 2

LITERATURE REVIEW

2.1 History of molecular imprinting

The first work on molecular imprinting was published in 1931 (Polyakov, 1931) on silica surfaces and continuously developed mainly on organic polymers. The development of silica imprinted remained silence after steady publications for 15 years. The main limitation during this period that it must used a water soluble template which must be fairly stable in acid. The new period of intensive development of molecular imprinted polymers was started in 1972 (Klotz & Takagishi, 1972; K. G. Wulff & Sarhan, 1972). They independently reported preparation of organic polymer with predetermined ligand selectivities. Briefly, the method is known as "controlled distance method", which involved copolymerization between D-glyceric-(*p*-vinylanilide)-2,3-O-*p*-vinylphenylboronate and divinylbenzene. Research and development related to MIP are kept on growing continuously until today with more than 4000 paper published ("MIP Database," 2010). The wide interests of scientific community to MIP technology are reflected with the number of publications as shown in Figure 2.1.