

REFERENCES

- Agriculture and Agri-Food Canada. (2014). "Market Overview: Malaysia" in *Market Access Secretariat Global Analysis Report*. Ottawa, Canada: Global Analysis Division.
- Alvarez, C., Reyes-Sosa, F. M., and Diez, B. (2016). Enzymatic hydrolysis of biomass from wood. *Microbial biotechnology*, 9(2), 149-156.
- Arduengo, P. M. (2012). Buffers for Biochemical Reactions. In A. P. M., *Protocols & Applications Guide* (pp. 151-154). America: Promega Corporation.
- Bajpai, P. (2016). *Pretreatment of Lignocellulosic Biomass for Biofuel Production*. Switzerland: Springer.
- Bensah, E. C., and Mensah, M. (2013). Chemical pretreatment methods for the production of cellulosic ethanol: Technologies and innovations. *International Journal of Chemical Engineering*, 1-21.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K. B., and Ramakrishnan, S. (2011). Chemical and physiochemical pretreatment of lignocellulosic biomass: A review. *Enzyme Research*, 10(1), 415-436.
- Bruice, P. Y. (2014). *Organic Chemistry*. London: Pearson Education Limited.
- Chang, J. S. (2011). Bioconversion and bioprocess technology for cleaner environment and better life. *Journal of the Taiwan Institute of Chemical Engineers*, 42, 3771-3774.
- Chi, G., Hu, S., Yang, Y., and Chen, T. (2012). Response surface methodology with prediction uncertainty: A multi-objective optimisation approach. *Chemical Engineering Research and Design*, 90, 1235-1244.
- Chidi, E. E., Oluwatisin, S. K., and Deborah, K. (2015). Microwave-alkaline assisted pretreatment of banana trunk for bioethanol production. *Journal of Energy and Power Engineering*, 9, 705-713.
- Cordeiro, N., Belgacem, M. N., Torres, I. C., and Moura, J. C. V. P. (2004). Chemical composition and pulping of banana pseudo-stems. *Industrial Crops and Products*, 19, 147-154.
- Economic Planning Unit, Malaysia. (2010). *Tenth Malaysia Plan*. Putrajaya, Malaysia: Prime Minister's Department.

- Filho, L. C. G., Fischer, G. A. A., Sellin, N., Marangoni, C., and Souza, O. (2013). Hydrolysis of banana tree pseudostem and second-generation ethanol production by *sacharomyces cerevisiae*. *Journal of Environment Science and Engineering*, 2, 65-69.
- Food and Agriculture Organization of the United Nations (FAO). (2014). *Banana Market Review and Banana Statistics 2012-2013*. Rome, Italy: Team on International Investment and Tropical Fruits.
- Food and Agriculture Organization of the United Nations (FAO). (2017). *Banana Market Review 2015-2016*. Rome, Italy: Team on International Investment and Tropical Fruits.
- Harmsen, P. F. H., Huijgen, W. J. J., Lopez, L. M. B., and Bakker, R. R. C. (2010). *Literature review of physical and chemical pretreatment processes for lignocellulosic biomass*. Netherlands: Wageningen University & Research Centre - Food & Biobased Research.
- Inoue, H., Yano, S., Endo, T., Sakaki, T., and Sawayama, S. (2008). Combining hot-compressed water and ball milling pretreatments to improve the efficiency of the enzymatic hydrolysis of eucalyptus. *Biotechnology for Biofuels*, 1(2), 1-9.
- Iroba, K. L., Tabil, L. G., Dumonceaux, T., and Baik, O. D. (2013). Effect of alkaline pretreatment on chemical composition of lignocellulosic biomass using radio frequency heating. *Biosystems Engineering*, 116(1), 385-398.
- Karimi, K., and Taherzadeh, J. (2016). A critical review on analysis in pretreatment of lignocelluloses: Degree of polymerization, adsorption/desorption, and accessibility. *Bioresource Technology*, 203(1), 348-356.
- Karimi, K., Shafiei, M., and Kumar, R. (2013). Chapter 3: Progress in Physical and Chemical Pretreatment of Lignocellulosic Biomass. In G. a. V. K., *Biofuel Technologies* (pp. 53-96). Germany: Springer-Verlag Berlin Heidelberg.
- Kashyap, M. C., Agrawal, Y. C., Ghosh, P. K., Jayas, D. S., Sarkar, B. C., and Singh, B. P. N. (2007). Enzymatic hydrolysis pretreatment to solvent extraction of soybrokens for enhanced oil availability and extractability. *Journal of Food Process Engineering*, 29(1), 664-674.
- Khuri, A. I. (2017). Response surface methodology and its applications in agricultural and food sciences. *Biometrics & Biostatistics International Journal*, 5(5), 128-149.
- Kumar, A. D., and Sharma, S. (2017). Recent updates on different methods of pretreatment of lignocellulosic feedstocks: A review. *Bioresources and Bioprocessing*, 4(7), 17-37.
- Kumar, P., Barrett, D. M., Delwiche, M. J., and Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial & Engineering Chemistry Research*, 48(8), 3713-3729.
- Lehto, J., and Alen, R. (2015). Alkaline pre-treatment of sftwood chips prior to delignification. *Journal of Wood Chemical Technology*, 35(2), 146-155.

- Lei, H., Cybulska, I., and Julson, J. (2013). Hydrothermal pretreatment of lignocellulosic biomass and kinetics. *Journal of Sustainable Bioenergy Systems*, 3(1), 250-259.
- Li, K., Fu, S., Zhan, Y., and Lucia, L. A. (2010). Analysis of the chemical composition and morphological structure of banana pseudo-stem. *Bioresources*, 5(2), 576-585.
- Lopez-Arenas, T., Rathi, P., Ramirez-Jimenez, E., and Sales-Cruz, M. (2010). Factors affecting the acid pretreatment of lignocellulosic biomass: Batch and continuous process. *Computer Aided Chemical Engineering*, 28(1), 979-984.
- Maurya, D. P., Singla, A., and Negi, S. (2015). An overview of key pretreatment processes for biological conversion of lignocellulosic biomass to bioethanol. *Biotechnology*, 5(5), 597-609.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31(3), 426-428.
- Mtui, G. (2009). Recent advances in pretreatment of lignocellulosic wastes and production of value added products. *African Journal of Biotechnology*, 8(8), 1398-1415.
- Myat, L. and Ryu, G. H. (2015). Pretreatments and factors affecting saccharification and fermentation for lignocellulosic ethanol production. *Cellulose Chemistry and Technology*, 50(2), 177-188.
- Olayiwola, O. M., Amahia, G. N., Adewara, A. A., and Chukwu, A. U. (2011). Application of response surface methodology for capturing optimum response in a longitudinal survey. *International Journal of Applied Science and Technology*, 1(5), 171-176.
- Paloheimo, M., Haarmann, T., Makinen, S., and Vehmaanpera. (2016). Production of Industrial Enzymes in *Trichoderma reesei*. In a. C. M. Schmoll, *Gene Expression System in Fungi: Advancements and Applications*, *Fungal Biology* (pp. 23-57). Switzerland: Springer International Publishing.
- Preethi, P., and Balakrishna, M. G. (2013). Physical and chemical properties of banana fibre extracted from commercial banana cultivars grown in Tamilnadu state. *Agrotecnology*, 23(11), 1-3.
- Rambo, M. K. D., Schmidt, F. L., and Ferreira, M. M. C. (2015). Analysis of the lignocellulosic components of biomass residues for biorefinery opportunities. *Talanta*, 144, 696-703.
- Singh, D. P., and Trivedi, R. K. (2013). Acid and alkaline pretreatment of lignocellulosic biomass to produce ethanol as biofuel. *International Journal of ChemTech Research*, 5(2), 727-734.
- Song, Z., Yang, G., Guo, Y., and Zhang, T. (2012). Comparison of two chemical pretreatment of rice straw for biogas production by anaerobic digestion. *Bioresources*, 7(3), 3223-3236.

- Teoh, Y. P., and Ooi, Z. X. (2016). Evaluation of unstructured kinetic models for the production of bioethanol from banana and pineapple wastes. *Bioresources*, 11(2), 4295-4305.
- Tutt, M., Kikas, T., and Olt, J. (2012). Comparison of different pretreatment methods on degradation of rye straw. *Engineering for Rural Development*, 24, 412-416.
- Wan Azelee, N. I., Md Jahim, J., Ismail, A. F., Mohamad Fuzi, S. F. Z., Rahman, R. A., Ghazali, N. F., and Md Illias, R. (2016). Enzymatic hydrolysis of pretreated Kenaf using a recombinant xylanase: Effects of reaction conditions for optimum hemicellulose hydrolysis. *American Journal of Agricultural and Biological Sciences*, 11(2), 54-66.
- Yabefa, J. A., Ocholi, Y., and Odubo, G. F. (2014). Effect of temperature and changes in medium pH on enzymatic hydrolysis of B(1-4) glycosidic bond in orange mesocarp. *Asian Journal of Plant Science and Research*, 4(2), 21-24.

© This item is protected by original copyright

APPENDIX A

PREPARATION OF 0.1 M CITRATE BUFFER SOLUTIONS

0.1 M citrate buffer was prepared from the mixture of 0.1 M citric acid monohydrate ($\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$) solution and 0.1 M trisodium citrate dihydrate ($\text{C}_6\text{H}_5\text{O}_7\text{Na}_3 \cdot 2\text{H}_2\text{O}$) solution. To prepare both $\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$ solution and $\text{C}_6\text{H}_5\text{O}_7\text{Na}_3 \cdot 2\text{H}_2\text{O}$ solution with molar concentration of 0.1 M, 21.01 grams of $\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$ and 29.41 grams of $\text{C}_6\text{H}_5\text{O}_7\text{Na}_3 \cdot 2\text{H}_2\text{O}$ were added and dissolved well with separated 1 L of distilled water respectively. The next step of 0.1 M citrate buffer with pH 5.0 was prepared according to Table A-1 by mixing 35.0 mL of 0.1 M $\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$ solution with 65.0 mL of 0.1 M $\text{C}_6\text{H}_5\text{O}_7\text{Na}_3 \cdot 2\text{H}_2\text{O}$ solution to achieve a total volume of 100 mL of citrate buffer solution.

Table A-1: Guideline for 0.1 M citrate buffer preparation within a specific pH range (Arduengo, 2012).

pH value	0.1 M $\text{C}_6\text{H}_8\text{O}_7 \cdot \text{H}_2\text{O}$ (mL)	0.1 M $\text{C}_6\text{H}_5\text{O}_7\text{Na}_3 \cdot 2\text{H}_2\text{O}$ (mL)
3.0	82.0	18.0
4.0	59.0	41.0
5.0	35.0	65.0
6.0	11.5	88.5

APPENDIX B

PREPARATION OF 3,5-DINITROSALICYLIC ACID (DNS) REAGENTS USING MILLER METHOD

3,5-dinitrosalicylic acid (DNS) reagents was prepared according to the miller method which function to determine the present of reducing sugar – glucose (Miller, 1959). 5 grams of DNS powders were weighed and then added into 100 mL of 2 M NaOH solution. Simultaneously, 150 grams of Na-K-tartrate which function as colour stabilizer were dissolved in 500 mL of distilled water. Next, both of the prepared mixture solutions were mixed thoroughly before stored as DNS reagent solution in labelled dark bottle and kept in refrigerator when the time not in usage.

APPENDIX C

PREPARATION OF GLUCOSE STANDARD CURVE FOR DATA ANALYSIS

A standard curve for glucose concentration analysis was constructed by initially preparing 0 g/L, 10 g/L, 20 g/L, 30 g/L, 40 g/L, 50 g/L, 60 g/L, 70 g/L, 80 g/L, 90 g/L, and 100 g/L of glucose solutions in distilled water and then 2 mL from each of the varying concentration of glucose solution was added with 2 mL of DNS reagents solution in test tube respectively. Further heating on the test tubes in water bath at 95 °C for 5 minutes was carried on and then the test tubes with mixture solutions were let to be cooled down with running tap water before addition of 8 mL of distilled water into the test tubes. Then, UV-vis spectrophotometer was used to determine the glucose concentration of samples at wavelength of 540 nm. Table C-1 and Figure C-1 respectively show the absorbance analysis and glucose standard curve with determined best fit line and linear regression with equation.

Table C-1: Absorbance analysis for glucose standard curve.

Glucose concentration (g/L)	Absorbance reading ($\Delta 540$ nm)	Standard deviation
0	0.0000	0.0000
10	0.0176	0.0002
20	0.0199	0.0006
30	0.0378	0.0007
40	0.0417	0.0002
50	0.0497	0.0003
60	0.0661	0.0019
70	0.0679	0.0024
80	0.0815	0.0002
90	0.0905	0.0001
100	0.1051	0.0006

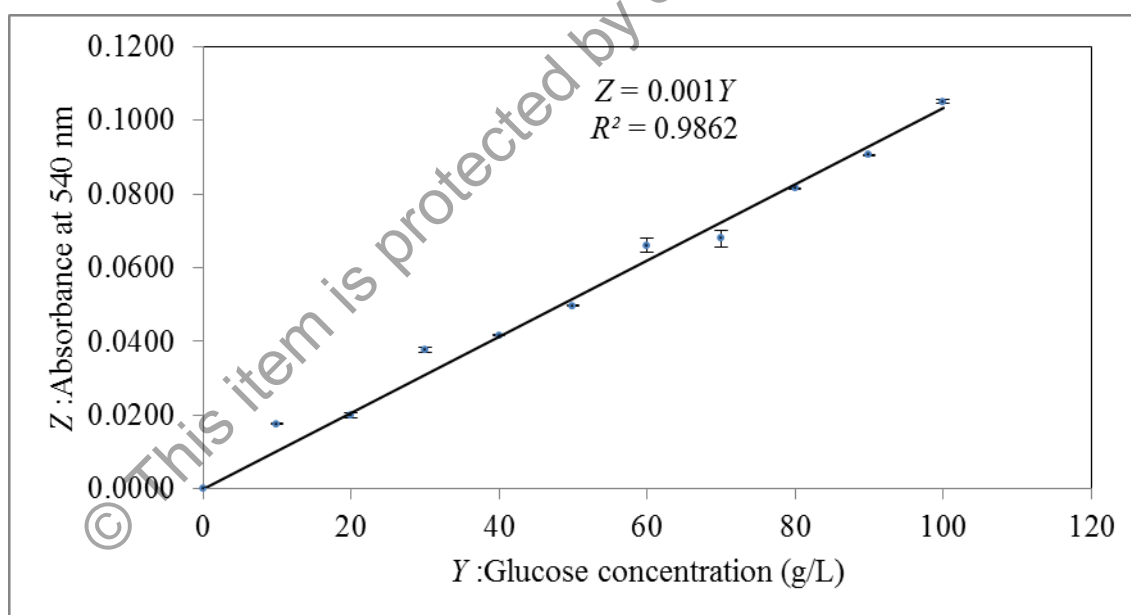


Figure C-1: Glucose standard curve.

APPENDIX D

RAW DATA FOR PRELIMINARY STUDIES: SELECTION OF BEST CHEMICAL PRETREATMENT ON BANANA TRUNK BIOMASS

The raw data of comparison studies between 2% (v/v) H_2SO_4 solution (acidic) and 2% (v/w) NaOH solution (alkaline) in banana trunk biomass pretreatment are listed in Table D-1.

Table D-1: Raw data of glucose production analysis for preliminary studies by using UV-vis spectrophotometer.

Chemical type	Absorbance reading ($\Delta 540$ nm)				Glucose concentration (g/L)	Standard deviation
	Reading 1	Reading 2	Reading 3	Average Reading		
2% (w/v) NaOH	0.0516	0.0518	0.0499	0.0511	51.10	1.0440
2% (v/v) H_2SO_4	0.0646	0.0646	0.0648	0.0647	64.47	0.1155

APPENDIX E

RAW DATA FOR ONE-FACTOR AT-A-TIME (OFAT) STUDIES

The raw data of One-Factor-at-A-Time (OFAT) studies for three influencing factors of substrate concentration, treatment duration, and treatment temperature are listed in Table E-2. In addition, Table E-1 shows the fixed parameters values for respectively OFAT studies parameters.

Table E-1: Fixed parameters values for respectively OFAT studies parameters.

OFAT studies	Fixed parameter	
Substrate concentration (% (w/v))	Treatment temperature	100 °C
	Treatment duration	30 minutes
Treatment duration (min)	Substrate concentration	10% (w/v)
	Treatment temperature	100 °C
Treatment temperature (°C)	Substrate concentration	10% (w/v)
	Treatment duration	30 minutes

Table E-2: Raw data of glucose production analysis for OFAT studies by using UV-vis spectrophotometer.

Substrate conc. (% (w/v))	Absorbance reading ($\Delta 540$ nm)				Glucose conc. (g/L)	Standard deviation
	Reading 1	Reading 2	Reading 3	Average Reading		
4	0.0290	0.0293	0.0291	0.0291	29.13	0.1528
6	0.0363	0.0362	0.0361	0.0362	36.20	0.1000
8	0.0518	0.0540	0.0542	0.0533	53.33	1.3317
10	0.0646	0.0646	0.0648	0.0647	64.67	0.1155
Treatment duration (min)	Absorbance reading ($\Delta 540$ nm)				Glucose conc. (g/L)	Standard deviation
	Reading 1	Reading 2	Reading 3	Average Reading		
10	0.0358	0.0358	0.0358	0.0358	35.80	0.0000
20	0.0513	0.0514	0.0514	0.0514	51.37	0.0577
30	0.0646	0.0646	0.0648	0.0647	64.67	0.1155
40	0.0551	0.0551	0.0551	0.0551	55.10	0.0000
Treatment temp. ($^{\circ}$ C)	Absorbance reading ($\Delta 540$ nm)				Glucose conc. (g/L)	Standard deviation
	Reading 1	Reading 2	Reading 3	Average Reading		
40	0.0122	0.0122	0.0122	0.0122	12.20	0.0000
60	0.0283	0.0283	0.0283	0.0283	28.30	0.0000
80	0.0356	0.0355	0.0352	0.0354	35.43	0.2082
100	0.0646	0.0646	0.0648	0.0647	64.67	0.1155

APPENDIX F

RAW DATA FOR OPTIMIZATION STUDIES: RESPONSE SURFACE METHODOLOGY (RSM)

The raw data of optimization studies of pretreatment process with consideration on the parameters of substrate concentration and treatment duration via RSM are listed in Table F-1.

Table F-1: Raw data for glucose production analysis for optimization studies by using UV-vis spectrophotometer.

Run	Substrate conc. (% (w/v))	Treatment duration (min)	Absorbance reading ($\Delta 540$ nm)				Glucose conc. (g/L)
			Reading 1	Reading 2	Reading 3	Average reading	
1	30.00	25.00	0.0817	0.0821	0.0817	0.0818	81.83
2	10.00	25.00	0.0539	0.0540	0.0538	0.0539	53.90
3	20.00	22.93	0.0755	0.0756	0.0753	0.0755	75.47
4	20.00	30.00	0.0840	0.0841	0.0840	0.0840	84.03
5	34.14	30.00	0.0814	0.0819	0.0815	0.0816	81.60
6	10.00	35.00	0.0668	0.0669	0.0668	0.0668	66.83
7	20.00	30.00	0.0844	0.0846	0.0844	0.0845	84.47
8	5.86	30.00	0.0310	0.0317	0.0312	0.0313	31.30
9	30.00	35.00	0.0889	0.0887	0.0888	0.0888	88.80
10	20.00	30.00	0.0840	0.0846	0.0846	0.0844	84.40
11	20.00	37.07	0.0996	0.0995	0.0996	0.0996	99.57
12	20.00	30.00	0.0843	0.0843	0.0843	0.0843	84.30
13	20.00	30.00	0.0847	0.0842	0.0842	0.0844	84.37

APPENDIX G

PHOTOGRAPH FOR RESEARCH PROJECT

The photos of experimental preparation and equipment used are taken and shown in Plate G-1 and G-2 respectively.

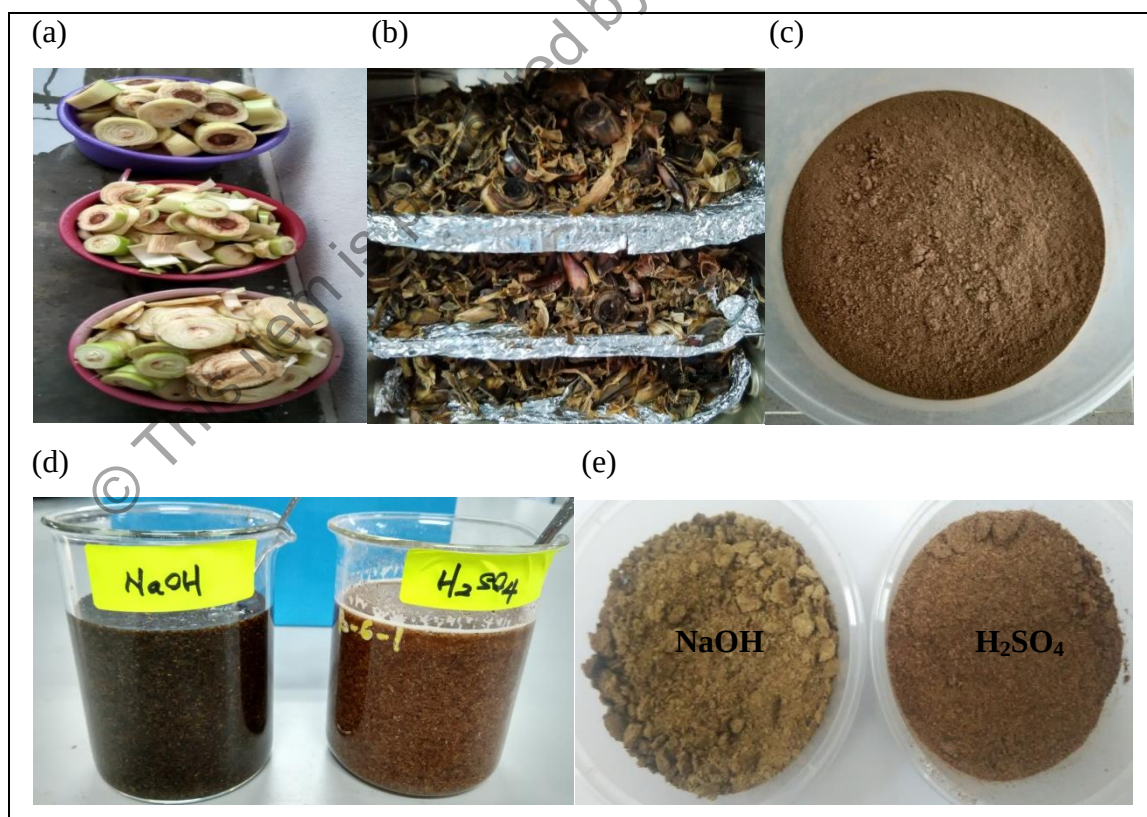


Plate G-1: Preparation of raw material (banana trunk) in the form of (a) sliced pieces, (b) dried pieces in oven, and (c) powder after grinded, (d) pretreated in chemicals, and (e) dried after pretreatment.



Plate G-2: Equipment used in research experiment including (a) oven, (b) grinder, (c) sieve shaker, (d) pH meter, (e) water bath, and (f) UV-vis spectrophotometer.